

This item is the archived peer-reviewed author-version of:

Dunaliella microalgae for nutritional protein : an undervalued asset

Reference:

Sui Yixing, Vlaeminck Siegfried.- Dunaliella microalgae for nutritional protein : an undervalued asset Trends in biotechnology : regular edition - ISSN 0167-7799 - London, Elsevier science london, 38:1(2020), p. 10-12 Full text (Publisher's DOI): https://doi.org/10.1016/J.TIBTECH.2019.07.011 To cite this reference: https://hdl.handle.net/10067/1649030151162165141

uantwerpen.be

Institutional repository IRUA

1	Dunaliella microalgae	for nutritional proteir	, an undervalued asset
---	-----------------------	-------------------------	------------------------

- 2 Yixing Sui and Siegfried E. Vlaeminck^{*}
- 3 LinkedIn Yixing Sui: <u>https://www.linkedin.com/in/yixingsui/</u>
- 4 Twitter Siegfried E. Vlaeminck: @SigifridoF
- 5 Research Group of Sustainable Energy, Air and Water Technology, Department of Bioscience
- 6 Engineering, University of Antwerp, Groenenborgerlaan 171, 2020 Antwerpen, Belgium
- 7 Website: <u>https://www.uantwerpen.be/en/research-groups/sustainable-energy/</u>
- 8 *Correspondence: <u>siegfried.vlaeminck@uantwerpen.be</u> (S.E. Vlaeminck)
- 9 Key words:
- 10 novel food; microbial protein; single-cell protein; essential amino acids; protein shift

11 Abstract:

- 12 β-carotene production with *Dunaliella* microalgae is established, yet their potential as protein
- 13 source for food and feed applications seems overlooked. The rich protein content and
- 14 nutritional tunability of *Dunaliella* make these algae intriguing sources of sustainable protein. It
- 15 is of societal interest to exploit these promising proteinaceous *Dunaliella* traits.

16 Dunaliella microalgae: Spotlighted β-carotene and undervalued protein

- 17 Microalgae are recognized as promising sources for diversified applications including food, feed
- 18 and high-value products [1]. Both science and industry have already focused their interests on
- 19 Dunaliella microalgae, especially for their unique feature of hyper carotenogenesis, producing

20	β -carotene as one of the first commercial high-value products from microalgae [2]. The large-
21	scale production of β -carotene from <i>Dunaliella</i> , as β -carotene extract or dried biomass, started
22	in the 1980s in Israel, Australia and USA, followed by other countries like India and China [2].
23	Dunaliella has been classified by the U.S. Food and Drug Administration (FDA) as food sources
24	with GRAS (Generally Regarded as Safe) status, and is mostly used for human and animal
25	nutrition, food coloring and cosmetics due to its pro-vitamin and anti-oxidant functions [1].
26	Among all commercial <i>Dunaliella</i> products, they are mostly extracts of β -carotene,
27	accompanied by powders in capsules or tablets. When possible, the residual of biomass allows
28	further usage as glycerol, protein, enzymes, fatty acids, vitamins etc.
29	Nevertheless, the societal challenge of food and protein scarcity are prompting a need for nove
30	protein sources to sustain the population growth, and microalgae present a resource-efficient
31	and ecological way of producing proteinaceous food and feed ingredients among proteins from
32	other microbes such as bacteria and yeast [3,4]. In this regard, the potential of Dunaliella
33	protein has been undervalued. From the 1950s until recently, although Dunaliella has been
34	noted for its proteinaceous traits, only scattered studies have been carried out. Even in the
35	established studies, patents and projects (e.g. European project D-Factory), Dunaliella protein
36	mostly comes as a by-product used for e.g. animal feeding after β -carotene extraction. These
37	facts leave Dunaliella protein rather poorly understood comparing with traditional microalgal
38	protein producers such as Chlorella and Spirulina. Therefore, it is of great interest to revisit and
39	further explore the potential usage of <i>Dunaliella</i> protein.

40 **Dunaliella** protein snapshot: Limitations of single-point analyses

The typical way to analyze protein in Dunaliella biomass is to assess its protein content and 41 42 protein composition from snapshot approaches, which are performed under specific conditions 43 and for specific growth phases (mostly in the stationary phase under standard cultivation 44 conditions) [4]. These approaches have shown that the dried biomass of Dunaliella consists of 45 50-80% protein [3,4]. More notably, the essential amino acid (EAA) composition of Dunaliella, 46 as indicated by essential amino acid index (EAAI), can reach superior protein quality for human 47 requirement following reference level set by Food and Agriculture Organization (FAO) [5] (Box 48 1). Compared with soybean and other protein-rich microalgae genera such as Spiruling and 49 *Chlorella, Dunaliella* also shows either equal or better quantity and quality of proteins [3]. 50 Differently from most other microalgae, Dunaliella microalgae lack rigid cellulosic cell wall, so 51 they are more easily digested by both humans and animals. Although the protein profile of 52 Dunaliella seems favorable, these snapshot approaches provide incomplete information and 53 neglect the potential variabilities introduced by external cultivation conditions.

54 Dunaliella protein tunability: Exploring the spectrum

The synthesis of biomass in microalgae strongly depends on nutrient level, salinity, pH, growth phase, and other conditions. Accordingly, studying the tunability of protein quantity and quality in *Dunaliella* can largely broaden the spectrum of snapshot analyses. For instance, nitrogen (N) is an essential element composing protein and amino acids (AA), so its availability can directly affect protein dynamics. High N availability can result in more protein accumulation in *Dunaliella*, provided that other nutrients are neither limiting nor overabundant [6]. Due to its halophilic characteristics, salinity also plays an important role. Around 10% sodium chloride

62 (NaCl) concentration has been reported to be optimal for both cell growth and protein quantity 63 [4,7]. Although pH level can be influential as well, it is mostly species dependent, with optima 64 ranging from pH 7.5 to pH 8.5 for maximum cell growth and protein quantity [4,7]. Higher light 65 intensity and longer illumination period seem to result in higher protein accumulation in 66 Dunaliella, yet they can also lower the light usage efficiency by microalgae, resulting in lower 67 protein yield [5,7,8]. Lastly, the growth phase of microalgae can affect the protein dynamics 68 significantly [4,5]. The exponential phase implies the generation of new cells, which is 69 accompanied by increased protein synthesis for cell reproduction [4,5]. The stationary phase, 70 however, implies a limiting condition which translates into slower cell growth and N 71 assimilation rates, resulting in lower protein quantity [4,5]. It is therefore common to find an 72 increase-decrease pattern of protein level throughout the growth phases [4,5]. 73 There exist large unidentified areas preventing an in-depth understanding on how AA dynamics 74 influence protein quality in Dunaliella. Only recently, one study demonstrated that EAA levels in 75 Dunaliella increase from the exponential towards the stationary phase, despite the increase-76 decrease pattern of protein quantity [5]. Another study shows a clear benefit of N limitation to 77 boost EAA production in Dunaliella despite reduced protein quantity [8]. Both studies indicate 78 the superior quality of *Dunaliella* protein by its essential amino acid index (EAAI) as high as 1.53 79 (Box 1). These studies show that further insights into dynamics of both protein quantity and 80 quality in *Dunaliella* are required to fully exploit its potential as a protein source.

81 Dunaliella protein in food and feed applications

82 From the 1980s until recently, very few studies were published on Dunaliella microalgae for 83 human and animal nutrition. The most pronounced benefits of food products containing dried 84 Dunaliella biomass, such as bread and pasta, is their enhanced nutritional properties, such as 85 increased proteins and minerals, and improved rheological properties [9,10]. Moreover, 86 Dunaliella biomass is better digested by rats (to mimic human digestion) compared with other 87 microalgae and casein, validating its "without rigid cell wall" advantage [11]. In husbandry, the 88 ovarian activity of goats is also improved when fed with *Dunaliella*-supplemented feed, 89 hastening the follicular development [12]. In aquaculture, living *Dunaliella* cells are mostly used 90 as feed ingredients, and sea urchins can efficiently incorporate Dunaliella protein into their 91 larvae [13]. These examples show that studies on the practical applications of *Dunaliella* protein 92 are indeed very scattered. Nevertheless, they provide positive prospects for favorable usage in 93 food and feed applications.

94 Future outlooks and concluding remarks

95 Single-product exploitation has been the primary focus of microalgal research for decades, even 96 though the remaining biomass after extraction of the main product is still ripe for further 97 valorization [14]. Some microalgae such as Chlorella and Spirulina producing lower-value 98 products e.g. protein and lipid, are also finding their higher-value paths towards pigments and 99 fatty acids including phycocyanin and linoleic acids. Nevertheless, these exclusive concepts 100 focusing on single product seems less beneficial comparing with a more inclusive strategy: co-101 production of nutritional compounds in microalgae [14]. As for Dunaliella, an emphasis on its 102 protein should not exclude its interesting β -carotene characteristic. Consequently, a new

103 approach to co-produce multiple products of interest (e.g. high quality protein and β -carotene 104 together) in Dunaliella biomass may be more advantageous [8]. Similarly, a co-production of 105 some functional proteins can further improve the quality of *Dunaliella* biomass. This biomass 106 can be subsequently used in several applications, bringing proteins with high quality and 107 digestibility, carotenoids with coloring, antioxidant and immune-stimulating functions together, 108 such as poultry feed, pet food, ornamental fish feed and ornamental bird feed [15] (Figure 1). 109 This approach could lead to win-win scenarios where microalgae production efficiency is 110 improved, resulting in products with higher values, and the relatively high production costs of 111 microalgae can be reduced.

112 The proteinaceous traits of *Dunaliella* present considerable potential for food and feed 113 applications. Nonetheless, the high-value β -carotene production from *Dunaliella* has largely 114 overshadowed its potential as a protein source. It is societally timely to focus on exploiting this 115 digestible high-quality protein, advance its co-production with β-carotene, and nutritionally 116 demonstrate the added value of the optimized products in multiple feed and food applications. 117 Furthermore, as concerned in all microalgal production and biorefinery domains, there still 118 exists various challenges on technological and social-economic aspects, encouraging 119 contributions from future endeavors.

120 Acknowledgement

This work was supported by the China Scholarship Council (File No. 201507650015) and the MIP
 i-Cleantech Flanders (Milieu-innovatieplatform; Environment innovation platform) project

Microbial Nutrients on Demand (MicroNOD). Dr. Michele Moretti from University of Antwerp isacknowledged for proofreading the manuscript.

125 Box 1. Essential amino acid index (EAAI) determines the protein quality for human food

Essential amino acids (EAA) cannot be synthesized by the human body, so humans rely on
external food supplies to provide them. EAAI scores can indicate the quality of a protein source
by comparing the ratios of EAA in the protein source to those required by the human body. The
following equation is used to calculate the EAAI score:

130
$$AAI = \sqrt[n]{\frac{aa_1}{AA_1} \times \frac{aa_2}{AA_2} \times \dots \times \frac{aa_n}{AA_n}}$$

where *aa_n* and *AA_n* are the EAA content over total protein (mg EAA/g protein) in the sample and
the FAO/WHO reference for human requirement, respectively [5]. EAAI values of ≥ 1, 0.95-1,
0.86-0.95, 0.75-0.86 and ≤ 0.75 correspond to superior, high, good, useful and inadequate
proteins quality, respectively [5]. For instance, based on the EAA composition of egg and
soybean, their EAAI scores are 1.65 and 1.34, respectively [3,5].

136 **Reference:**

141

3

Milledge, J.J. (2011) Commercial application of microalgae other than as biofuels: a brief
 review. *Rev. Environ. Sci. Bio/Technology* 10, 31–41
 Borowitzka, M.A. (2013) High-value products from microalgae-their development and
 commercialisation. *J. Appl. Phycol.* 25, 743–756

Becker, E.W. (2007) Micro-algae as a source of protein. *Biotechnol. Adv.* 25, 207–210

142	4	Sui, Y. and Vlaeminck, S.E. (2019) Effects of salinity, pH and growth phase on the protein
143		productivity by Dunaliella salina. J. Chem. Technol. Biotechnol. 94, 1032–1040
144	5	Sui, Y. et al. (2019) Light regime and growth phase affect the microalgal production of
145		protein quantity and quality with Dunaliella salina. Bioresour. Technol. 275, 145–152
146	6	Uriarte, I. et al. (1993) Cell characteristics and biochemical composition of Dunaliella
147		primolecta Butcher conditioned at different concentrations of dissolved nitrogen. J. Appl.
148		Phycol. 5, 447–453
149	7	Tavallaie, S. et al. (2015) Comparative studies of β -carotene and protein production from
150		Dunaliella salina isolated from lake Hoze-soltan, Iran. J. Aquat. Food Prod. Technol. 24,
151		79–90
152	8	Sui, Y. et al. (2019) Enhancement of co-production of nutritional protein and carotenoids
153		in Dunaliella salina using a two-phase cultivation assisted by nitrogen level and light
154		intensity. Bioresour. Technol. 287, 121398
155	9	Finney, K.F. et al. (1984) Use of algae Dunaliella as a protein supplement in bread. Cereal
156		Chem. 61, 402–406
157	10	El-Baz, F.K. et al. (2017) Microalgae Dunaliella salina for use as food supplement to
158		improve pasta quality. Int. J. Pharm. Sci. Rev. Res. 46, 45–51
159	11	Herrero, C. et al. (1993) Nutritional properties of four marine microalgae for albino rats.
160		J. Appl. Phycol. 5, 573–580
161	12	Senosy, W. et al. (2017) Effects of feeding green microalgae on ovarian activity,

162		reproductive hormones and metabolic parameters of Boer goats in arid subtropics.
163		Theriogenology 96, 16–22
164	13	Qi, S. et al. (2018) The effects of 3 different microalgae species on the growth,
165		metamorphosis and MYP gene expression of two sea urchins, Strongylocentrotus
166		intermedius and S. nudus. Aquaculture 492, 123–131
167	14	Gifuni, I. et al. (2019) Current Bottlenecks and Challenges of the Microalgal Biorefinery.
168		Trends Biotechnol. 37, 242–252
169	15	Amaya, E. et al. (2014) Carotenoids in Animal Nutrition, Fefana Publication.

170