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1 **Comparison of typical nitrite oxidizing bacteria suppression strategies**  
2 **and the effect on nitrous oxide emissions in a biofilm reactor**

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10 **Abstract:**

11 In mainstream partial nitrification/anammox (PN/A), suppression of nitrite oxidizing  
12 bacteria (NOB) and mitigation of N<sub>2</sub>O emissions are two essential operational goals.  
13 The N<sub>2</sub>O emissions linked to three typical NOB suppression strategies were tested in  
14 a covered rotating biological contactor biofilm system at 21°C: (i) low dissolved oxygen  
15 (DO) concentrations, and treatments with (ii) free ammonia (FA), and (iii) free nitrous  
16 acids (FNA). Low emerged DO levels effectively minimized NOB activity and decreased  
17 N<sub>2</sub>O emissions, but NOB adaptation appeared after 200 days of operation. Further  
18 NOB suppression was successfully achieved by periodic (3 hours per week) treatments  
19 with FA (29.3 ± 2.6 mg NH<sub>3</sub>-N L<sup>-1</sup>) or FNA (3.1 ± 0.3 mg HNO<sub>2</sub>-N L<sup>-1</sup>). FA treatment,  
20 however, promoted N<sub>2</sub>O emissions, while FNA did not affect these. Hence, biofilm  
21 PN/A should be operated at relatively low DO levels with periodic FNA treatment to  
22 maximize nitrogen removal efficiency while avoiding high greenhouse gas emissions.

23 **Keywords:** Deammonification; Nitrous oxide; Carbon footprint; Sewage; Biological

24 nutrient removal; Nitrification

## 25 **1. Introduction**

26 Partial nitritation/anammox (PN/A), a cost-effective and energy-efficient nitrogen  
27 removal process based on anoxic ammonium oxidation (anammox), has attracted  
28 increasing attention (Agrawal et al., 2018). Several successful PN/A applications have  
29 been reported in the side stream (sludge line) (Lackner et al., 2014). For mainstream  
30 PN/A, i.e. application to the sewage water line, it is more challenging to achieve long-  
31 term stability than for the side stream process, which is mainly attributed to the lower  
32 temperature in temperate climates (10-15°C versus >30°C) and lower nitrogen  
33 concentrations making nitrite oxidizing bacteria (NOB) suppression difficult (Laureni  
34 et al., 2016; Peng et al., 2020; Zhu et al., 2022). The key goal is that the activities of  
35 the ‘functional’ anoxic- ammonium oxidizing bacteria (AnAOB) and aerobic  
36 ammonium oxidizing bacteria (AerAOB) can be maintained while unwanted NOB are  
37 effectively inhibited. Till now, strategies that relied on the physiological characteristics  
38 of these three groups of bacteria have been tested. Specifically for NOB, the observed  
39 growth rate ( $dX_{NOB}/dt$ ) was shown in Eq. 1 (Laureni et al., 2019; Wang et al., 2021),  
40 indicating that NOB suppression can be obtained using three main strategies, e.g.,  
41 reducing bacterial growth rate ( $\mu_{NOB,max}$ ), promoting biomass decay rate ( $b_{NOB}$ ), and  
42 increasing NOB’s washout (sludge retention time (SRT) decrease).

$$43 \quad \frac{dX_{NOB}}{X_{NOB} \cdot dt} = \left( \mu_{NOB,max} - b_{NOB} - \frac{1}{SRT} \right) \quad (\text{Eq. 1})$$

44 To reduce  $\mu_{NOB,max}$ , low dissolved oxygen (DO) concentrations (Hausherr et al., 2022;  
45 Ma et al., 2011) or intermittent aeration (Gu et al., 2022; Peng et al., 2020) are

46 commonly used in the reaction system because NOB has a higher half-saturation  
47 constant of  $O_2$  compared to AerAOB, which means that NOB lose more activity as the  
48 DO level decreases (Blackburne et al., 2008). To promote  $b_{NOB}$ , free ammonia (FA, the  
49 un-ionized form of ammonium) treatment (Wang et al., 2017), free nitrous acid (FNA,  
50 the protonated form of nitrite) treatment (Peng et al., 2020), or alternating FA/FNA  
51 shocks (Duan et al., 2019) have been successfully applied. The low FA and FNA levels  
52 in the mainstream (due to the low nitrogen concentrations and temperature) limit the  
53 possibility to directly suppress NOB, but this could be tackled by return-sludge  
54 treatment (high FA and FNA conditions could be achieved in the side stream) (Peng et  
55 al., 2020). To increase the value of  $1/SRT$ , 'SRT control' was applied to remove NOB  
56 (e.g., flocs) while maintaining AnAOB (e.g., biofilm) (Agrawal et al., 2018). An  
57 integrated fixed-film activated sludge reactor (IFAS), where AnAOB grow on carriers  
58 (long-biofilm) and NOB commonly on flocs (short-floc), or a two-stage PN/A process,  
59 where the 'partial nitrification' and 'anammox' processes were separated into two  
60 reactors, has been used (Peng et al., 2020; Seuntjens et al., 2018).

61 The strategies described above are usually accompanied by high ammonium  
62 (potentially during FA treatment) and high nitrite concentrations (potentially during  
63 FNA treatment), and the strategies can be potentially powerful inducers of nitrous  
64 oxide ( $N_2O$ ) production and emission (Laureni et al, 2016; Peng et al., 2014). That led  
65 to variations in  $N_2O$  emissions (3.0 – 6.4% of the influent nitrogen loading) from  
66 mainstream PN/A systems (Ali et al., 2016; Connan et al., 2018; De Clippeleir et al.,  
67 2013).  $N_2O$  is a potent greenhouse gas (~265-fold stronger in global warming potential  
68 than carbon dioxide ( $CO_2$ )) and a strong ozone-depleting substance (Peng et al., 2020;  
69 Ravishankara et al., 2009). There are two main routes for AerAOB to produce  $N_2O$ , i.e.,

70 the nitrifier denitrification route ( $\text{NO}_2^- \rightarrow \text{NO} \rightarrow \text{N}_2\text{O}$ ) (Kampschreur et al., 2008) and  
71 the  $\text{NH}_2\text{OH}$  oxidation route (a side product of the incomplete oxidation process:  $\text{NH}_4^+$   
72  $\rightarrow \text{NH}_2\text{OH} \rightarrow \text{NO}_2^-$ ) (Peng et al., 2014). Nitrite and DO levels are thus critical factors  
73 affecting  $\text{N}_2\text{O}$  production. In addition, there is a third  $\text{N}_2\text{O}$  production route when  
74 organic carbon is present (Ma et al., 2017), i.e., heterotrophic denitrification (anoxic  
75 reduction of  $\text{NO}_2^-$  or  $\text{NO}_3^- \rightarrow \text{N}_2\text{O}$ ).

76 Only by studying the influences of different NOB suppression strategies on  $\text{N}_2\text{O}$   
77 emissions, the optimal strategy that suppresses NOB activity while limiting  $\text{N}_2\text{O}$   
78 emissions and carbon footprint, can be identified. Although multiple NOB suppression  
79 strategies have been employed in various studies, as well as the linked  $\text{N}_2\text{O}$  emissions,  
80 there is a lack of research that compares these distinct methods within a single system.  
81 This is crucial to ensure a fair comparison, as the results can be influenced by factors  
82 such as microbial community composition and types of sludge. Furthermore, there is  
83 also a dearth of knowledge regarding the dynamic characteristics of  $\text{N}_2\text{O}$  emissions  
84 during the treatment process and between different treatment approaches. Thus,  
85 there is a need to further investigate and understand these aspects.

86 The present research's overall objective was to test the effectiveness of typical NOB  
87 suppression strategies and quantify the accompanying  $\text{N}_2\text{O}$  emissions. To achieve that,  
88 the  $\text{N}_2\text{O}$  emissions linked to three common NOB strategies, i.e., low DO levels control,  
89 periodic FA treatment, and periodic FNA treatment, were characterized.

## 90 **2. Materials and methods**

### 91 **2.1. Rotating biological contactor (RBC)**

92 An RBC with mature PN/A biofilm was operated at  $21 \pm 0.6^\circ\text{C}$  for 550 days (Figure 1).

93 Detailed information on this reactor was presented by Van Tendeloo et al. (2021). The  
94 discs' submersion level was fixed at 50%, corresponding to an effective volume of 51  
95 L. With a disc rotation speed of 1.8 rpm, a consecutive exposure of 17 s emerged and  
96 submerged condition was achieved. The RBC was covered by an airtight overhead  
97 cover, creating a controlled headspace. The O<sub>2</sub> concentration in this headspace (i.e.,  
98 the emerged DO level) was controlled by the inlet flow rates of N<sub>2</sub> and compressed air  
99 to the headspace. Except for days 401 to 450 when the effect of different emerged  
100 DO levels was assessed, the conditions were fixed at 0.60 mg O<sub>2</sub> L<sup>-1</sup> (the values were  
101 ranging between 0.19 – 1.84 mg O<sub>2</sub> L<sup>-1</sup> during days 401 to 450).

102 The emerged DO levels meant the O<sub>2</sub> concentration in the liquid around the biofilm,  
103 which was estimated from the gas phase measurement in the present research (the  
104 corresponding O<sub>2</sub> level, which could be dissolved in the liquid film, linked to O<sub>2</sub> in the  
105 gas phase). Microorganisms always take up the O<sub>2</sub> from the liquid film. According to  
106 previous research, 85–89% of the O<sub>2</sub> input was directly absorbed during the air  
107 exposure of the discs (air phase) (Courstens et al., 2014). The off-gas was actively  
108 pumped out by a gas analyzer at 330 L h<sup>-1</sup>, which equalled to the inlet gas flow rate  
109 (the total inlet flow rate of N<sub>2</sub> and compressed air).

110 The synthetic mainstream wastewater was made from tap water supplemented  
111 with (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> (47.6 ± 2.3 mg N L<sup>-1</sup>), NaHCO<sub>3</sub> (5 mg HCO<sub>3</sub><sup>-</sup> mg<sup>-1</sup> N), KH<sub>2</sub>PO<sub>4</sub> (7.5 mg P L<sup>-1</sup>  
112 <sup>1</sup>), and trace elements solutions A/B (0.01 ml L<sup>-1</sup>) (de Graaf et al., 1995). The influent  
113 flow rate was kept at ~120 L d<sup>-1</sup> in a continuous mode (Watson-Marlow 323, United  
114 Kingdom), resulting in a nitrogen loading rate of 105.2 ± 5.2 mg N L<sup>-1</sup> d<sup>-1</sup>.

## 115 **2.2 Overall Experimental Plan**

116 The experiment was divided into four phases.

### 117 **2.2.1 Phase – I: Long-term operation at low emerged DO level**

118 In Phase – I (from days 0 to 400), the long-term stability of mainstream PN/A under  
119 low emerged DO level was studied. Because of the high initial NOB activity, the  
120 emerged DO level was reduced from  $0.80 \text{ O}_2 \text{ L}^{-1}$  to  $0.60 \text{ mg O}_2 \text{ L}^{-1}$  on day 26 to suppress  
121 NOB activity. This value ( $0.60 \text{ mg O}_2 \text{ L}^{-1}$ ) was selected since the reactivation of NOB  
122 was observed in the biofilm after the DO increased to values higher than  $0.65 \text{ mg O}_2$   
123  $\text{L}^{-1}$  (Wang et al., 2018). Afterward, a period of up to 374 days was used to verify the  
124 stability of NOB inhibition by a low emerged DO level.

### 125 **2.2.2 Phase – II: Effect of different emerged DO levels on the N<sub>2</sub>O emission**

126 In Phase – II (from days 401 to 450), the effect of emerged DO levels on the  
127 performance of the PN/A process and N<sub>2</sub>O emission was investigated. The DO set  
128 point was manipulated within the range of  $0.2\text{-}1.8 \text{ mg O}_2 \text{ L}^{-1}$ , with increments of  $0.4$   
129  $\text{mg O}_2 \text{ L}^{-1}$  (average experimental value:  $0.19, 0.60, 0.91, 1.43, \text{ and } 1.84 \text{ mg O}_2 \text{ L}^{-1}$ ). This  
130 adjustment was made by modifying the influent flow rates of N<sub>2</sub> (ranging from  $8.8 \text{ L h}^{-1}$   
131  $^1$  to  $11.4 \text{ L h}^{-1}$ ) and compressed air (ranging from  $0.2 \text{ L h}^{-1}$  to  $2.8 \text{ L h}^{-1}$ ) into the  
132 headspace. Each DO level was tested for only 2 days to avoid a strong (irreversible)  
133 increase in NOB activity at higher emerged DO conditions. Subsequently, the emerged  
134 DO level was returned to  $\sim 0.60 \text{ mg O}_2 \text{ L}^{-1}$ . When the  $\text{NH}_4^+$  and  $\text{NO}_3^-$  conversion rates  
135 were recovered to the benchmark, the next emerged DO level was tested.

### 136 **2.2.3 Phase – III: Effect of FA treatment on the N<sub>2</sub>O emission**

137 In Phase – III (from days 451 to 497), FA was used as a stressor for NOB suppression.

138 In total, three FA treatments (3 hours per treatment) were performed with a  
139 treatment interval of 7 days (days 453, 460, and 467). The FA treatment condition  
140 ( $29.3 \pm 2.6 \text{ mg NH}_3\text{-N L}^{-1}$  achieved at a pH of  $\sim 8.0$ , temperature of  $\sim 21^\circ\text{C}$ , and  
141 ammonium concentration of  $\sim 735 \text{ mg N L}^{-1}$ ) was chosen based on the previous  
142 research (Van Tendeloo et al., 2021). After the RBC recovered from the FA treatment,  
143 the isolated effect of high pH (8.0, 3h) and high ammonium concentration ( $735 \text{ mg N}$   
144  $\text{L}^{-1}$ , 3h) shock was tested on days 487 – 490 and days 480 – 484, respectively, to verify  
145 the effect of these two factors on the  $\text{N}_2\text{O}$  emissions. The emerged DO level was stable  
146 at  $0.60 \text{ mg O}_2 \text{ L}^{-1}$  throughout the phase.

#### 147 **2.2.4 Phase – IV: Effect of FNA treatment on the $\text{N}_2\text{O}$ emission**

148 The NOB activity was positively correlated to the FNA treatment frequency (Duan et  
149 al., 2018; Peng et al., 2020). However, the higher treatment frequency and FNA levels  
150 would lead to a significant reduction in AerAOB and AnAOB activity. The FA and FNA  
151 levels selected in this research were based on the previous research based on realistic  
152 values in STP. In Phase – IV (from days 498 to 550), three FNA biofilm treatments were  
153 applied. The FNA treatments lasted for 3 hours and were performed with a treatment  
154 interval of 7 days (days 498, 505, and 512). The FNA treatment condition ( $3.1 \pm 0.3 \text{ mg}$   
155  $\text{HNO}_2\text{-N L}^{-1}$  achieved at a pH of  $\sim 6.0$ , temperature of  $\sim 21^\circ\text{C}$ , and nitrite concentration  
156 of  $\sim 1205 \text{ mg N L}^{-1}$ ) was chosen according to Peng et al. (2020). After the RBC recovered  
157 from the FNA treatment, the isolated effect of low pH (6.0, 3h) and high nitrite  
158 concentration ( $1205 \text{ mg N L}^{-1}$ , 3h) shock was tested on days 531 – 536 and 540 – 547,  
159 respectively, to verify the effect of these two factors on the  $\text{N}_2\text{O}$  emissions. During the  
160 whole period, the emerged DO level was again stable at  $0.60 \text{ mg O}_2 \text{ L}^{-1}$ .

161 **2.3 Online N<sub>2</sub>O emission monitoring**

162 The off-gas was dehumidified with a gas cooler (Bühler Technologies, Germany) and  
163 analyzed with an online N<sub>2</sub>O gas analyzer (Emerson Rosemount CT5800 Quantum  
164 Cascade Laser Gas Analyzer, United States, United States) with a range of 0 – 500 ppm  
165 (the lowest detection (LOD) was 2 ppm). The data was logged every 30 seconds. Zero  
166 and span calibration were accomplished by calibrating the N<sub>2</sub>O analyzer  
167 measurements against N<sub>2</sub> of instrument gas purity and N<sub>2</sub>O reference gas. The  
168 sampling flow rate was set at 330 L h<sup>-1</sup> by a vacuum pump (KNF Laboport, the  
169 Netherlands) which equalled the inlet flow rate (N<sub>2</sub> and compressed air) to maintain  
170 the gas balance.

171 The N<sub>2</sub>O emission factor (%) was calculated according to the following equations:

172 
$$\text{N}_2\text{O emission factor} = \text{N}_2\text{O emitted} \div \text{NH}_4^+ \text{ converted} \quad (\text{Eq. 2})$$

173 
$$\text{N}_2\text{O emitted} = \sum (C_{\text{N}_2\text{O gas}} \times Q_{\text{sampling pump}} \times \Delta t) \quad (\text{Eq. 3})$$

174 
$$\text{NH}_4^+ \text{ converted} = \text{NH}_4^+ \text{ conversion rate} \times Q_{\text{influent}} \times \Delta t \quad (\text{Eq. 4})$$

175 where N<sub>2</sub>O emitted represents the released N<sub>2</sub>O over a certain amount of time [mg  
176 N<sub>2</sub>O-N]; NH<sub>4</sub><sup>+</sup> converted represents the converted NH<sub>4</sub><sup>+</sup> over a certain amount of time  
177 [mg NH<sub>4</sub><sup>+</sup>-N];  $C_{\text{N}_2\text{O gas}}$  represents the point N<sub>2</sub>O concentration in the off-gas [mg N<sub>2</sub>O-  
178 N L<sup>-1</sup>];  $Q_{\text{sampling pump}}$  represents the flow rate of the gas sampling pump [330 L h<sup>-1</sup>];  $\Delta t$   
179 represents the time interval by which the off-gas N<sub>2</sub>O concentration was recorded [h];  
180 NH<sub>4</sub><sup>+</sup> conversion rate is measured by daily sampling [mg N L<sup>-1</sup> d<sup>-1</sup>];  $Q_{\text{influent}}$  represents  
181 the flow rate of the influent (L h<sup>-1</sup>). The N<sub>2</sub>O concentration in the off-gas in ppm was  
182 converted to mg N<sub>2</sub>O-N L<sup>-1</sup> based on the volume occupied by 1 mole of an ideal gas at  
183 standard temperature and pressure (0°C and 101.3 kPa), which is 22.4 L and corrected  
184 for temperature of the gas sample (24°C).

185 The N<sub>2</sub>O emission factor is calculated based on the average N<sub>2</sub>O emitted within one  
186 day (continuous N<sub>2</sub>O measurement (per 30 seconds) and daily water samples (for NH<sub>4</sub><sup>+</sup>  
187 converted measurement)). The data were averaged over the entire period (from the  
188 moment the DO value changed to the moment it returned to 0.6 mg O<sub>2</sub> L<sup>-1</sup>) in the tests  
189 at different DO levels. The background values for the indoor N<sub>2</sub>O were also measured,  
190 which was almost negligible (the average N<sub>2</sub>O emission factor was <0.2%).

#### 191 **2.4. Analytical procedures**

192 Liquid samples were taken periodically from the influent and effluent to determine  
193 the PN/A performance. After filtering through a 0.2 µm syringe filter (CHROMAFIL Xtra  
194 PVDF, Germany) and storing at 4°C, ammonium, nitrite, and nitrate concentrations  
195 were measured with a San<sup>++</sup> Automated Wet Chemistry Analyzer (SKALAR, the  
196 Netherlands) and Ion Chromatograph with a Metrosep A Supp 5- 150/4.0 column  
197 (Metrohm – Eco IC, Switzerland). The pH and DO values were measured using  
198 handheld meters (Hach HQ30d, United States). The microbial community analysis  
199 followed the procedure described in Zhu et al. (2022): 16S rRNA gene amplicon  
200 sequence variants (ASVs) were determined using the V4 region. The data have been  
201 deposited with links to BioProject accession number PRJNA797575 in the NCBI  
202 BioProject database.

203 The p-value obtained from the analysis of variance (ANOVA) was compared with a  
204 significance level of 0.05 to assess the level of variation. A p-value below 0.05 was  
205 considered acceptable evidence of a significant difference. The FA and FNA  
206 concentrations were calculated according to Anthonisen et al. (1976).

#### 207 **2.5 Calculations: The relative NO<sub>2</sub><sup>-</sup> consumption by AnAOB and NOB**

208 It was assumed that the decrease in total nitrogen was converted to N<sub>2</sub> by the  
 209 anammox (the feed did not contain COD in the present study). During the anammox  
 210 process, the stoichiometric ratio of removed NO<sub>2</sub><sup>-</sup>/removed NH<sub>4</sub><sup>+</sup> and produced NO<sub>3</sub><sup>-</sup>  
 211 /removed NH<sub>4</sub><sup>+</sup> are 1.23 and 0.21, respectively (the average of the value reported by  
 212 Strous et al. (1998) and Lotti et al. (2014)). The percentage of NO<sub>2</sub><sup>-</sup> consumed by  
 213 AnAOB (P(AnAOB)), NOB (P(NOBS)), and excess (P(excess)) are shown in Eq. 5 to Eq. 10  
 214 (TN presents the total nitrogen).

$$215 \quad P(\text{AnAOB}) = \frac{\text{NO}_2^- \text{ consumed by AnAOB}}{\text{Influent NO}_2^- + \text{NO}_2^- \text{ produced by AerAOB}} \quad (\text{Eq. 5})$$

$$216 \quad P(\text{excess}) = \frac{\text{Remained NO}_2^-}{\text{Influent NO}_2^- + \text{NO}_2^- \text{ produced by AerAOB}} \quad (\text{Eq. 6})$$

$$217 \quad P(\text{NOB}) = 1 - P(\text{AnAOB}) - P(\text{excess}) \quad (\text{Eq. 7})$$

$$218 \quad \text{NO}_2^- \text{ produced by AerAOB} = \text{NH}_4^+ \text{ removed} - \text{NH}_4^+ \text{ consumed by AnAOB} \quad (\text{Eq. 8})$$

$$219 \quad \text{NO}_2^- \text{ consumed by AnAOB} = \frac{1.23}{1 + 1.23 - 0.21} \times \text{TN removed} \quad (\text{Eq. 9})$$

$$220 \quad \text{NH}_4^+ \text{ consumed by AnAOB} = \frac{1}{1 + 1.23 - 0.21} \times \text{TN removed} \quad (\text{Eq. 10})$$

## 221 **3. Results and discussion**

### 222 **3.1 Long-term operation at low emerged DO levels**

#### 223 **3.1.1 Decrease the emerged DO level suppressed NOB**

224 Long-term stability is essential for successfully implementing mainstream PN/A at full  
 225 scale. To assess the potential for this, the performance of the RBC was evaluated for  
 226 400 days at a low emerged DO level. Initially, the emerged DO level was set at 0.80 mg  
 227 O<sub>2</sub> L<sup>-1</sup>. This, however, boosted NOB activity, resulting in a gradual increase in produced  
 228 NO<sub>3</sub><sup>-</sup> and a NO<sub>2</sub><sup>-</sup> consumption by NOB of more than 50% (Figure 2). To suppress NOB,

229 the emerged DO level was decreased to  $0.60 \pm 0.02 \text{ mg O}_2 \text{ L}^{-1}$  by changing the ratio  
230 between the inlet flow rates of  $\text{N}_2$  (from  $8.8 \text{ L h}^{-1}$  to  $11.4 \text{ L h}^{-1}$ ) and compressed air  
231 (from  $0.2 \text{ L h}^{-1}$  to  $2.8 \text{ L h}^{-1}$ ) on day 26. Between days 26 and 200, the  $\text{NO}_3^-$  production  
232 rate (from  $33.8$  to  $5.8 \text{ mg N L}^{-1} \text{ d}^{-1}$ ) and the percentage of  $\text{NO}_2^-$ -N consumed by NOB  
233 (from about 52% to about 0%) gradually decreased (Figure 2), indicating that NOB  
234 activity was effectively reduced by the decrease of emerged DO levels. This is in line  
235 with Zhu et al. (2023) and Van Tendeloo et al. (2021) who demonstrated that strict  $\text{O}_2$   
236 control could achieve complete NOB suppression.

237 The  $\text{NH}_4^+$  removal rate remained stable between days 26 and 200 ( $71.7 \pm 5.7 \text{ mg}$   
238  $\text{NH}_4^+\text{-N L}^{-1} \text{ d}^{-1}$ ) (Figure 2A), indicating that decreasing the DO level from  $0.8 \text{ mg O}_2 \text{ L}^{-1}$   
239 to  $0.60 \text{ mg O}_2 \text{ L}^{-1}$  could effectively suppress NOB activity as well as maintain AerAOB  
240 and AnAOB activities. In previous studies, DO levels in a biofilm PN/A system were  
241 controlled below  $0.20 \text{ mg O}_2 \text{ L}^{-1}$  (to maintain enough AnAOB) to suppress NOB. Still,  
242 this strategy lowered AerAOB activity, limiting the overall nitrogen removal rate  
243 (Wang et al., 2021). The biofilm in the RBC faced an intermittent aeration mode,  
244 switching between anoxic (i.e., submerged DO level of  $0 \text{ mg O}_2 \text{ L}^{-1}$ ) and low DO (i.e.,  
245 emerged DO level of  $0.60 \text{ mg O}_2 \text{ L}^{-1}$ ) every 17 seconds (i.e., transient anoxia). This  
246 might balance the regular supply of  $\text{O}_2$  for AerAOB to produce  $\text{NO}_2^-$ , followed by the  
247 removal of the produced  $\text{NO}_2^-$  during the anoxic period by AnAOB while suppressing  
248 NOB activity.

249 After 200 days of operation at an emerged DO level of  $0.60 \text{ mg O}_2 \text{ L}^{-1}$ , the  $\text{NO}_2^-$   
250 consumption by NOB started to increase (e.g., 0% at day 204 versus 30% at day 392)  
251 (Figure 2). This might be due to the adaptation of the suppressed NOB to the low DO  
252 conditions (i.e., NOB became more efficient in competing with AerAOB for  $\text{O}_2$  and with

253 AnAOB for NO<sub>2</sub><sup>-</sup>) (Cao et al., 2018). Microbial community analysis revealed that the  
254 enrichment in K-strategists *Nitrospira* (from 0.8% before the DO change on day 26 to  
255 4.7% on day 402), a NOB genus with a higher affinity to O<sub>2</sub> than others (e.g.,  
256 *Nitrobacter* and *Nitrotoga*) (Wang et al., 2021), in the PN/A biofilm (see  
257 supplementary material). The enrichment of *Nitrospira* would eventually result in the  
258 failure of the nitrite shunt, which led to the deterioration of the PN/A system (Duan  
259 et al., 2019). Cao et al. (2018) also revealed the enrichment of *Nitrospira* during the  
260 process of NOB adaptation to low DO level. At the same time, NOB are also likely to  
261 maintain structural and genetic integrity by repairing and minimizing damage to the  
262 cellular infrastructure at low DO levels (Duan et al., 2019). The adaptation of NOB to  
263 low DO levels after the long-term operation (> 200 days) has also been reported by  
264 Cao et al. (2018). Even though reducing the emerged DO level from 0.8 mg O<sub>2</sub> L<sup>-1</sup> to  
265 0.6 mg O<sub>2</sub> L<sup>-1</sup> could inhibit NOB activity, the adaptation of NOB to the low DO  
266 conditions occurred in the long-term run. Thus, to achieve a successful and stable  
267 PN/A system, the combined treatment built upon the low DO control to suppress NOB  
268 is worthy of further investigation.

### 269 **3.1.2 Decreasing the emerged DO levels reduced the N<sub>2</sub>O emission**

270 To investigate the effect of DO control on N<sub>2</sub>O emissions from the RBC, the emerged  
271 DO levels varied from 0.19 to 1.84 mg O<sub>2</sub> L<sup>-1</sup> between days 401 and 450. At an emerged  
272 DO level of 0.60 mg O<sub>2</sub> L<sup>-1</sup>, the average N<sub>2</sub>O emission factor was 1.92 ± 0.47% which is  
273 within the typical range for a PN/A system (0.1 – 2.4% reviewed by Ali et al. (2016)).  
274 As shown in Figure 3, the N<sub>2</sub>O emission factor increased with the increase of the  
275 emerged DO levels. At a DO of 0.19 mg O<sub>2</sub> L<sup>-1</sup>, the N<sub>2</sub>O emission factor was 1.59 ±

276 0.05%, while at a DO of 1.84 mg O<sub>2</sub> L<sup>-1</sup>, it was 2.48 ± 0.20%. One-way ANOVA analysis  
277 showed a significant positive correlation ( $R^2 = 0.9994$ ,  $p < 0.05$ ) between the emerged  
278 DO level and N<sub>2</sub>O emission factor at the DO level lower than 0.91 mg O<sub>2</sub> L<sup>-1</sup>. At higher  
279 emerged DO levels, the correlation was no longer significant ( $R^2 = 0.9224$ ,  $p > 0.05$ ).

280 Since the nitrifier denitrification N<sub>2</sub>O production route is linked to the nitrite  
281 concentration (Ma et al., 2017), it possibly had only limited contribution to the N<sub>2</sub>O  
282 emission in this study because of the low nitrite concentration (0.6 ± 0.8 mg N L<sup>-1</sup>) in  
283 the system. Furthermore, heterotrophic denitrification was likely negligible since it  
284 could only dominate when heterotrophy was stimulated in the presence of COD  
285 (whereas the feed in the present study did not contain COD) (Ma et al., 2017).  
286 Therefore, N<sub>2</sub>O emission might be mainly attributed to the NH<sub>2</sub>OH oxidation pathway.  
287 The positive correlation between the DO level and the N<sub>2</sub>O emission via the NH<sub>2</sub>OH  
288 oxidation route was also demonstrated in the nitrifier community of previous research  
289 (Domingo-Félez et al., 2014; Wunderlin et al., 2013). Hence, a low DO level is  
290 recommended to decrease N<sub>2</sub>O emissions from PN/A systems.

291 The emission of N<sub>2</sub>O was dynamic (the average values were used when calculating  
292 the N<sub>2</sub>O emission factor) and was, amongst others, influenced by changes in operating  
293 conditions in the reactor. A shift in emerged DO level caused an immediate change in  
294 the emitted N<sub>2</sub>O rate. Even when the DO level was not changed, the N<sub>2</sub>O  
295 concentration in the headspace varied. This might be caused by accumulation of NO<sub>2</sub><sup>-</sup>,  
296 light, small differences in temperature (± 1°C) and microbial respiration, variable gas  
297 pressure, or carbon fixation rate (Lotti et al., 2014). These findings are in line with Qiu  
298 et al. (2021), who also found that the observed rate of N<sub>2</sub>O production was dynamic  
299 even at a constant nitrification activity.

## 300           **3.2 FA treatment on PN/A biofilm**

### 301           **3.2.1 NOB suppression was further improved by FA treatment**

302 Long-term operation can trigger adaptation of NOB to low emerged DO conditions  
303 (Section 3.1). In Phase – III (from days 451 to 497), FA treatment ( $29.3 \pm 2.6$  mg  $\text{NH}_3$ -  
304 N  $\text{L}^{-1}$ ) was therefore implemented in combination with the low emerged DO level  
305 ( $\sim 0.60$  mg  $\text{O}_2$   $\text{L}^{-1}$ ) to suppress NOB activity (Figure 4). Before the first FA treatment, an  
306 average of  $17.5 \pm 9.3\%$  of the  $\text{NO}_2^-$  was consumed by NOB. This value decreased to 0%  
307 on day 455 (two days after the first FA treatment), indicating that the activity of NOB  
308 was completely suppressed by FA treatment. The following two FA treatments kept  
309 the NOB activity low between days 453 and 473 (e.g., 1.9% on day 470). The  
310 effectiveness of FA treatment is consistent with Van Tendeloo et al. (2021). After the  
311 last FA treatment, the inhibited NOB activity was gradually restored (e.g., 22.7% of the  
312  $\text{NO}_2^-$  was consumed by the NOB on day 480, 12 days after the last FA treatment),  
313 which is also in line with Duan et al. (2019) who reported that stopping FA treatment  
314 disrupted the established  $\text{NO}_2^-$  shunt. In conclusion, FA treatment successfully  
315 suppressed low DO-adapted NOB activity, but periodic treatments (3 hours per week)  
316 were required to maintain the suppression.

317       There is no unified conclusion regarding the mechanism of FA on NOB suppression.  
318 One possibility is that FA directly exhibits inhibitory effects on enzymes involved in  
319 nitrite oxidation reactions in NOB (Vadivelu et al., 2006). This hindrance makes  
320 electron transfer during the respiration process more challenging for NOB, ultimately  
321 reducing their activity. Another possibility suggests that FA may have an inhibitory  
322 effect on the synthesis of adenosine triphosphate (ATP) in NOB, which significantly

323 hampers their anabolism (Yuan, 2007). The precise mechanisms and interactions  
324 between FA and NOB are still subjects of ongoing research, and further studies are  
325 necessary to comprehensively understand the inhibition mechanisms of FA on various  
326 NOB species.

327 During FA treatment, the  $\text{NH}_4^+$  removal rate decreased ( $68.2 \pm 6.1$  versus  $76.9 \pm 4.6$   
328  $\text{mg N L}^{-1} \text{d}^{-1}$ ), indicating that AerAOB activity was also influenced by the FA treatment  
329 ( $30 \text{ mg NH}_3\text{-N L}^{-1}$ ). The residual nitrite concentration was increased ( $9.1 \pm 6.7\%$  versus  
330  $<5\%$  before FA treatment), suggesting that the inhibitory effect of FA treatment on  
331 NOB might be stronger than that on AerAOB. A more selective inhibition on NOB  
332 guaranteed a stable operation of the PN/A system. Seuntjens et al. (2018) also used a  
333 similar FA level ( $30 \text{ mg NH}_3\text{-N L}^{-1}$ , 3 hours) to suppress NOB and reported that FA  
334 treatment as the sole strategy could not inhibit NOB (increase in relative activity ratio  
335 between AerAOB and NOB). This suggests that the combination of FA treatment with  
336 a low DO level control is critical to obtain NOB suppression.

### 337 **3.2.2 FA treatment increased the $\text{N}_2\text{O}$ emission**

338  $\text{N}_2\text{O}$  concentrations in the air-phase of the RBC system were measured to assess the  
339 effect of FA treatment on the  $\text{N}_2\text{O}$  emissions. Before the FA treatment (benchmark,  
340 days 350 to 400), the  $\text{N}_2\text{O}$  emission factor was  $1.65 \pm 0.10\%$ . It rapidly increased to  
341  $2.33 \pm 0.36\%$ ,  $2.40 \pm 0.45\%$ , and  $2.15 \pm 0.30\%$  after each FA treatment (days 453, 460,  
342 and 467), respectively (Figure 5), revealing that FA treatment promoted  $\text{N}_2\text{O}$  emission  
343 from the RBC system. After the final FA treatment, the  $\text{N}_2\text{O}$  emission factor gradually  
344 decreased back to the benchmark within a week. To distinguish the effect of the FA  
345 treatment from the influence of the higher ammonium concentration and pH level,

346 separated high pH ( $\sim 8.0$ , 3h) and high ammonium concentration ( $\sim 735 \text{ mg N L}^{-1}$ , 3h)  
347 shocks were tested on days 488 – 490 and days 480 – 484. Results showed that the  
348 high pH level did not affect the  $\text{N}_2\text{O}$  emission factor ( $1.68 \pm 0.08\%$ ), whereas the  $\text{NH}_4^+$   
349 shock promoted the emission of  $\text{N}_2\text{O}$  ( $2.27 \pm 0.13\%$  versus  $1.65 \pm 0.10\%$  from the  
350 benchmark), suggesting that the high ammonium concentration might be the main  
351 contributor to the high  $\text{N}_2\text{O}$  emission of FA treatment. Kampschreur et al. (2009) also  
352 reported that the  $\text{N}_2\text{O}$  emission increased with increasing ammonium concentration  
353 in a full-scale STP.

354 DO and  $\text{NO}_2^-$  levels are two other essential factors affecting  $\text{N}_2\text{O}$  production by  
355 AerAOB (Peng et al., 2014; Wunderlin et al., 2013). The DO levels were always constant  
356 during the tests that the residual  $\text{NO}_2^-$  levels were likely the main influencing factor  
357 since it was positively correlated with the  $\text{N}_2\text{O}$  (Peng et al., 2015). After the FA  
358 treatment, due to the residual  $\text{NO}_2^-$  increase ( $10.7 \pm 6.7\%$  versus  $4.2 \pm 1.7\%$ ), the  
359 nitrifier denitrification  $\text{N}_2\text{O}$  production pathway might also play a role. The nitrifier  
360 denitrification route is nitrite-sensitive because the expression of copper-containing  
361  $\text{NO}_2^-$  reductase (NirK) is regulated by a nitrite-sensitive transcription repressor protein  
362 (Beaumont et al., 2004). The lower residual  $\text{NO}_2^-$  after the third FA treatment  
363 corresponded to the lower  $\text{N}_2\text{O}$  emission factor, which confirmed the role of residual  
364  $\text{NO}_2^-$ . Due to the operation being kept constant after the FA treatment (e.g., influent  
365 ammonium concentrations and the emerged DO levels), the  $\text{N}_2\text{O}$  produced through  
366 the  $\text{NH}_2\text{OH}$  oxidation route was likely the same as before. Thus, the  $\text{N}_2\text{O}$  emission  
367 increased after the FA treatment might be attributed to the rate increase of nitrifier  
368 denitrification route in the present research.

369 The  $\text{N}_2\text{O}$  emission was also dynamic during the FA treatments. At the beginning of

370 the FA treatment, the N<sub>2</sub>O concentration in the air phase rapidly increased (e.g., from  
371 1.03 to 4.23 ppm within 30 minutes during the first FA treatment). Subsequently, the  
372 N<sub>2</sub>O production gradually decreased (e.g., from 4.23 to 2.31 ppm). A peak was  
373 observed after 3 hours of FA treatment, followed by a decrease in N<sub>2</sub>O production.  
374 Even after 8 hours, the N<sub>2</sub>O emission was still higher than the N<sub>2</sub>O emission before the  
375 FA treatment. The high residual ammonium concentration might have increased the  
376 NH<sub>4</sub><sup>+</sup> oxidation rate, which probably yielded more intermediates (e.g., HNO, a  
377 byproduct during the oxidation process: NH<sub>2</sub>OH → N<sub>2</sub>O) (Law et al., 2012). The NH<sub>4</sub><sup>+</sup>  
378 shock caused a rapid increase in N<sub>2</sub>O concentration (e.g., from 4.02 to 24.06 ppm in  
379 the first shock). Although the value gradually decreased to 16.15 ppm after 3 hours,  
380 probably due to the decrease in ammonium concentration, it was still more than 10  
381 times higher than the baseline. At high NH<sub>4</sub><sup>+</sup> levels, not only the NH<sub>2</sub>OH oxidation  
382 route in AerAOB was promoted, but also N<sub>2</sub>O emissions from AnAOB were reported  
383 to be enhanced (Jin et al., 2016).

384 The high pH (8.0) shock had a limited influence on N<sub>2</sub>O emission (the drop during  
385 the shock can be attributed to the cessation of influent). The effect of pH on N<sub>2</sub>O  
386 emissions was likely directly related to the influence on the bacterial enzymes and  
387 nitrogen compounds rather than corresponding to a shift in bacterial communities  
388 (not long enough for a change). A pH range of 6.0 – 8.0 is not expected to affect the  
389 ammonia oxidation rate, and was therefore not expected to affect N<sub>2</sub>O emissions. It  
390 was agreed by Ribera-Guardia and Pijuan (2017) that the N<sub>2</sub>O emission was not  
391 influenced by a pH decrease (8.0 → 6.5).

### 392 **3.3 FNA treatment on PN/A biofilm**

393

### 3.3.1 NOB suppression was further improved by FNA treatment

394 After stopping the FA treatment, the percentage of  $\text{NO}_2^-$  consumed by NOB increased  
395 to  $13.5 \pm 5.6 \%$ , showing the recovery of NOB activity. In Phase – IV (from days 498 –  
396 550), FNA treatment ( $3.1 \pm 0.3 \text{ mg HNO}_2\text{-N L}^{-1}$ ) was applied in combination with low  
397 emerged DO levels ( $\sim 0.60 \text{ mg O}_2 \text{ L}^{-1}$ ) (Figure 4) to suppress NOB activity. The FNA  
398 treatment rapidly inhibited the NOB activity, decreasing the percentage of nitrite  
399 consumed by NOB (0%). Wang et al. (2014) also found that FNA treatment ( $0.24 - 1.35$   
400  $\text{mg HNO}_2\text{-N L}^{-1}$ ) had a significantly higher biocidal effect on NOB than AerAOB. After  
401 the last FNA treatment on day 512, the inhibited NOB activity was suppressed for the  
402 following 15 days (e.g., only 4.1% of  $\text{NO}_2^-$  was consumed by the NOB on day 527). That  
403 was different from FA treatment in which the activity of NOB was rapidly recovered  
404 after the treatment ( $\sim 6$  days versus  $\sim 15$  days). To conclude, low-DO adapted NOB  
405 were successfully suppressed by FNA treatment, but periodic treatment is also  
406 required to sustain the suppression. Wang et al. (2016a) also reported that low DO  
407 conditions ( $0.30 - 0.80 \text{ mg O}_2 \text{ L}^{-1}$ ) combined with the FNA treatment successfully  
408 suppressed NOB in the mainstream PN/A system. This finding is contradicting results  
409 of Van Tendeloo et al. (2021) who showed that FNA treatments with similar  
410 concentrations failed to suppress NOB activity, which might be attributed to the  
411 difference in reactor setup (RBC without overhead cover), operating conditions  
412 (higher emerged and submerged DO levels), and the community composition (e.g.,  
413 dominant NOB genus was *Nitrotoga* versus *Nitrospira* in the present research).

414 FNA has the ability to enhance the permeability of the cell membrane, facilitating  
415 the entry of nitrite into the cells. This change in the intracellular environment can  
416 disrupt the functioning of enzymes involved in electron transfer and proton

417 translocation processes (Philips et al., 2002). Additionally, FNA can inhibit the enzymes  
418 responsible for electron transfer and proton translocation in NOB, subsequently  
419 impeding ATP synthesis and affecting reactions catalyzed by ATPases. As a result, the  
420 activity of NOB is diminished (Frison et al., 2013).

421 After the FNA treatment, the  $\text{NH}_4^+$  removal rate did not differ from the value before  
422 treatment ( $77.9 \pm 5.5$  versus  $76.9 \pm 4.6 \text{ mg N L}^{-1} \text{ d}^{-1}$ ). In addition, almost no residual  
423  $\text{NO}_2^-$  ( $0.8 \pm 0.6 \text{ mg N L}^{-1}$ ) was present in the RBC system. Both results indicated that  
424 AnAOB and AerAOB activities were not affected by FNA treatment. In contrast, Peng  
425 et al. (2020) reported that FNA treatment exerted an inhibitory impact on both  
426 AerAOB and NOB, while the inhibition on NOB was stronger than AerAOB. This  
427 difference might be attributed to the higher stability of the biofilm structure in RBC.

### 428 **3.3.2 FNA treatment did not affect $\text{N}_2\text{O}$ emission**

429 The  $\text{N}_2\text{O}$  emission factor after FNA treatment (based on the low-DO conditions) was  
430 also evaluated. The values were  $1.73 \pm 0.27\%$ ,  $1.64 \pm 0.24\%$ , and  $1.64 \pm 0.34\%$  after  
431 each FNA treatment, respectively (Figure 6). There was no difference compared to the  
432 benchmark ( $1.72 \pm 0.16\%$ ) before the first FNA treatment. That differed from the FA  
433 treatment, which promoted the  $\text{N}_2\text{O}$  emissions in the RBC system. To distinguish the  
434 effect of FNA treatment on the  $\text{N}_2\text{O}$  emission from the effect of a nitrite concentration  
435 and pH level, low pH value ( $\sim 6.0$ , 3h) and high nitrite concentration ( $\sim 1205 \text{ mg N L}^{-1}$ ,  
436 3h) shocks were tested on days 531 – 536 and days 540 – 547. Both the low pH ( $1.68$   
437  $\pm 0.23\%$ ) and high nitrite concentration ( $1.70 \pm 0.16\%$ ) did not affect the  $\text{N}_2\text{O}$  emission  
438 factor ( $1.65 \pm 0.10\%$  in the benchmark), corresponding to the fact that FNA treatment  
439 does not affect  $\text{N}_2\text{O}$  emissions.

440 Even if the high  $\text{NO}_2^-$  during the FNA treatment (3h) promoted the  $\text{N}_2\text{O}$  emission,  
441 the nitrite concentration was reduced to  $< 1.0 \text{ mg N L}^{-1}$  within two hours. After the  
442 FNA treatment, there is no change in the residual  $\text{NO}_2^-$  ( $2.6 \pm 2.2\%$  versus  $4.2 \pm 1.7\%$ ),  
443 suggesting the nitrifier denitrification  $\text{N}_2\text{O}$  production pathway likely still did not play  
444 a role, which is again in contrast to the results after the FA treatment (increased  
445 residual  $\text{NO}_2^-$  promoted  $\text{N}_2\text{O}$  emission). In addition, the operation was also constant  
446 with that before the treatment, indicating similar  $\text{N}_2\text{O}$  emitted from the PN/A system.

447 As mentioned above, the  $\text{N}_2\text{O}$  emissions were always dynamic, including during the  
448 FNA treatment process. During the FNA treatment, the  $\text{N}_2\text{O}$  production gradually  
449 decreased (e.g., from 4.15 to 3.27 ppm within 3 hours in the first FNA treatment). Due  
450 to the presence of high  $\text{NO}_2^-$ , the nitrifier denitrification was likely the primary  $\text{N}_2\text{O}$   
451 production pathway during FNA treatment (Peng et al., 2015). According to previous  
452 research, the nitrifier denitrification process is stimulated at moderate  $\text{NO}_2^-$  range (0  
453 –  $50 \text{ mg N L}^{-1}$ ) (Peng et al., 2015), whereas inhibition appeared at high concentrations  
454 ( $500 - 1000 \text{ mg N L}^{-1}$ ) (Law et al., 2013). The  $\text{NO}_2^-$  level used in the present research  
455 ( $1205 \text{ mg N L}^{-1}$ ) was already beyond the reported suppression range, which might  
456 change the detoxification mechanism of NirK (copper-containing nitrite reductase),  
457 altering the stimulation threshold of  $\text{NO}_2^-$  to  $\text{N}_2\text{O}$  emissions (Wang et al., 2016b). That  
458 could explain the limited influence of FNA treatment on the  $\text{N}_2\text{O}$  emission. After 3  
459 hours of FNA treatment, the production gradually returned to the benchmark. Due to  
460 the sensitivity of the  $\text{N}_2\text{O}$  emission to high nitrite concentration being relatively large  
461 (Tallec et al., 2006), the  $\text{N}_2\text{O}$  production rapidly increased at the beginning of the  $\text{NO}_2^-$   
462 shock (e.g., from 3.91 to 6.33 ppm within 10 minutes in the first  $\text{NO}_2^-$  shock).  
463 Subsequently, the  $\text{N}_2\text{O}$  production gradually increased (e.g., from 6.33 to 10.91 ppm).

464 That might be explained by the previous findings (Ma et al., 2017) that the nitrite  
465 concentration increase could promote the AerAOB denitrification rate (effectively  
466 reducing  $\text{NO}_2^-$  to  $\text{N}_2\text{O}$ ). Regarding the low pH (6.0) shock, its impact on  $\text{N}_2\text{O}$  emissions  
467 was also limited, similar to the high pH (8.0) shock.

### 468 **3.4 Implication and outlook**

469 Regarding the suppression of NOB activity in mainstream PN/A systems, scientists  
470 have conducted different types of research, but studies of  $\text{N}_2\text{O}$  emissions from this  
471 process are still lacking. Strategies to suppress NOB are always accompanied by  
472 changes in operating parameters (e.g., pH, ammonium or nitrite concentration,  
473 temperature, etc.), leading to dynamic conditions that probably promote  $\text{N}_2\text{O}$   
474 emissions (Kampschreur et al., 2009). Decreasing emerged DO level could suppress  
475 NOB activity in the short run while mitigating  $\text{N}_2\text{O}$  emissions. However, NOB  
476 adaptation occurred at low DO levels during long-term operation (>200 days).  
477 Therefore, in addition to controlling low DO levels, FNA treatment should be  
478 performed periodically, since it stabilizes  $\text{N}_2\text{O}$  emissions while suppressing NOB  
479 activity. The findings in the present study are a beneficial supplement to the  
480 application of PN/A in the mainstream. While some breakthroughs have been made,  
481 there are still several issues that still merit further research. First, the correlation  
482 between periodical treatment and the levels of FNA is still unclear. Understanding the  
483 relationship between the two can provide a basis for selecting the most cost-effective  
484 frequency and level of FNA treatment. Second, the stability of the successful strategy  
485 proved in the present research, periodical FNA treatment combined with low DO  
486 control, should also be tested in the long-term run (> 200 days).

487 RBC is a type of biofilm-only system, which has higher biomass density and  
488 operational stability than the other reactors, such as suspended cell reactors (Soares  
489 et al., 2019). Yet, RBC is probably not a preferred system to apply in mainstream  
490 anammox. But if use them, several units that are coupled in series or parallel can be  
491 applied to treat the large flow wastewater. It would be at least technologically feasible  
492 or durable to cover them, mainly to protect the biofilm from weather events and lower  
493 the O<sub>2</sub> levels in the headspace (even gas control is economically not feasible). Even  
494 though the biomass growth configuration was different from the floccular and  
495 granular system (attached growth versus suspended growth), the findings of the  
496 present research are generic, and it can be extrapolated to several types of biofilm-  
497 based systems (e.g., integrated fixed-film activated sludge (IFAS), moving bed biofilm  
498 reactor (MBBR), etc.) (Van Tendeloo et al., 2021). At the level of N<sub>2</sub>O emissions, the  
499 type of system or the type of growth may not matter too much since it is mainly  
500 determined by the nitrite accumulation, O<sub>2</sub> level, and operational parameters.

#### 501 **4. Conclusion**

502 A balance between NOB suppression and N<sub>2</sub>O mitigation will be key to achieving a  
503 sustainable mainstream PN/A. Decrease emerged DO level suppressed NOB activity  
504 while mitigating N<sub>2</sub>O emissions. Yet, an adaptation of NOB occurred after 200 days  
505 operation. Periodic FA or FNA treatment further suppressed the low DO-adapted NOB.  
506 Characterizing the N<sub>2</sub>O emissions linked, FNA treatment did not affect (~1.67% versus  
507 1.72% of removed NH<sub>4</sub><sup>+</sup>-N) and was, hence, more advantageous than FA treatment  
508 (~2.29% versus ~1.65% of removed NH<sub>4</sub><sup>+</sup>-N). Thus, low DO conditions with periodic  
509 FNA treatment could effectively suppress NOB activity while emitting less N<sub>2</sub>O than

510 FA treatment.

511 E-supplementary data for this work can be found in e-version of this paper online.

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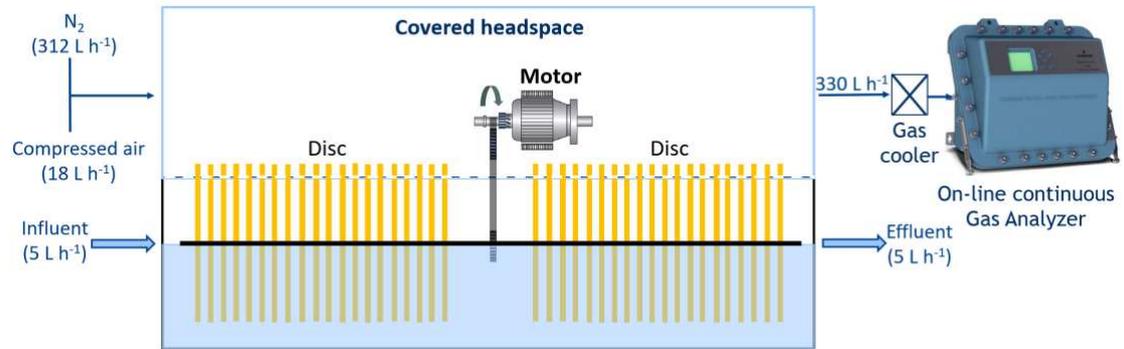
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676 **Table and Figure Captions**

677 **Figure 1**

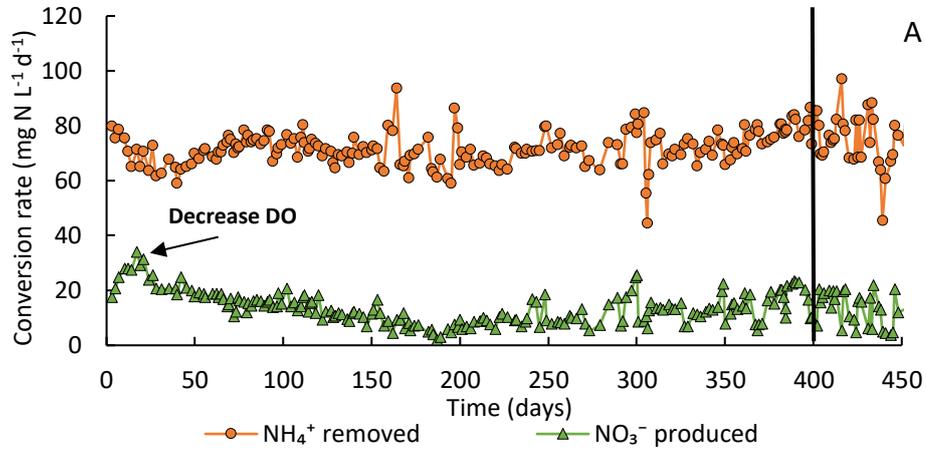


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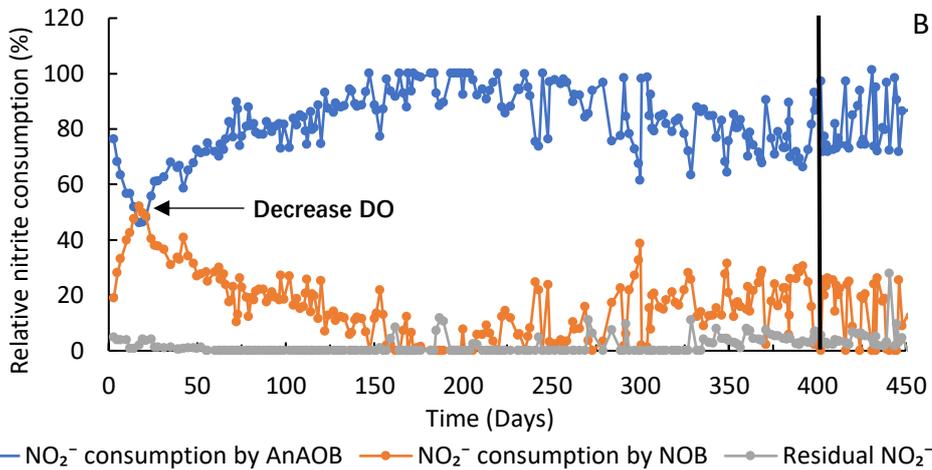
679 **Figure 1. Schematic of the rotating biological contactor (with overhead cover).**

680 **Figure 2**

Phase	Phase – I	Phase – II
Emerged DO	~ 0.60 mg O <sub>2</sub> L <sup>-1</sup> (~ 0.8 mg O <sub>2</sub> L <sup>-1</sup> before days 26)	0.19 – 1.84
Eff. NH <sub>4</sub> <sup>+</sup>	13.9 ± 3.5 mg N L <sup>-1</sup>	12.9 ± 5.3



681



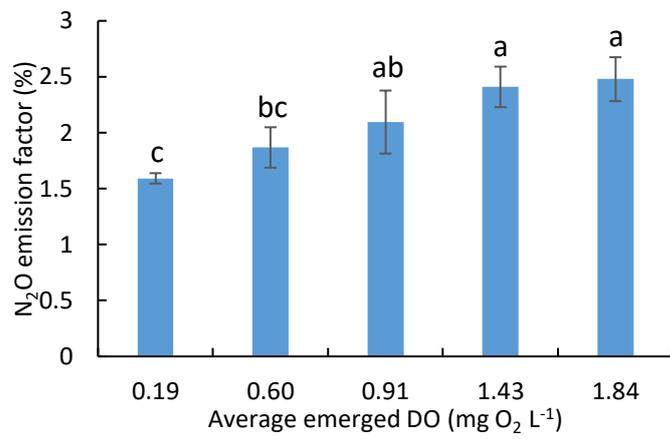
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683 **Figure 2. Reactor performance in Phase – I (days 0 to 400) and Phase – II (days 401**

684 **to 450). A, volumetric NH<sub>4</sub><sup>+</sup> removal and NO<sub>3</sub><sup>-</sup> production rate; B, relative NO<sub>2</sub><sup>-</sup>**

685 **consumption by AnAOB and NOB, and residual NO<sub>2</sub><sup>-</sup> (in the effluent).**

686 **Figure 3**



687

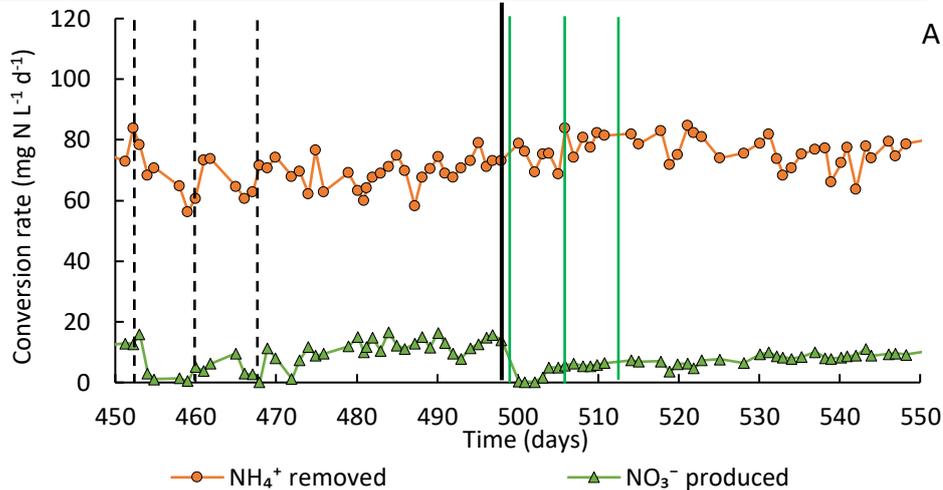
688 **Figure 3. The average N<sub>2</sub>O emission factor under different emerged DO levels.** The

689 error bars depict the standard deviations (n = 3). Significant differences are marked

690 with a letter ( $p < 0.05$ ).

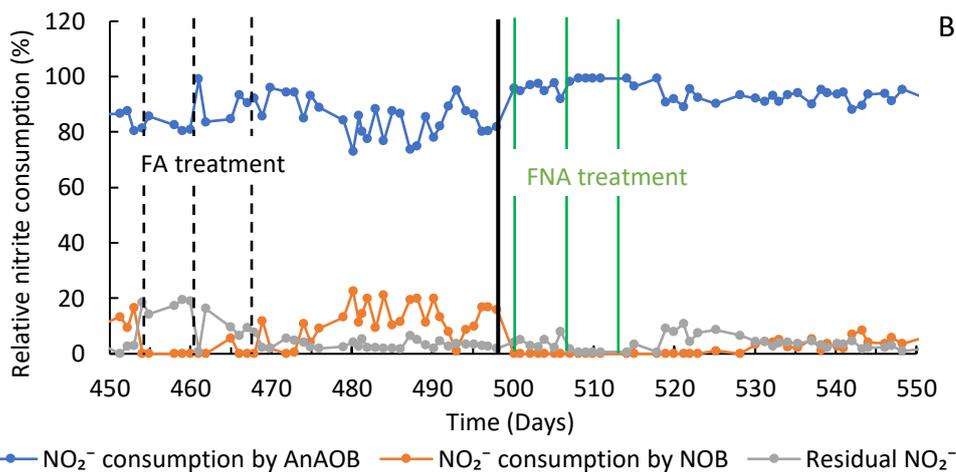
691 **Figure 4**

Phase	Phase – III	Phase – IV
DO level	~ 0.60 mg O <sub>2</sub> L <sup>-1</sup>	
Eff. NH <sub>4</sub> <sup>+</sup>	13.3 ± 3.8 mg N L <sup>-1</sup>	
FA	30 mg NH <sub>3</sub> -N L <sup>-1</sup>	/
FNA	/	3.0 mg HNO <sub>2</sub> -N L <sup>-1</sup>



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693



694

695 **Figure 4. Reactor performance in Phase – III (days 451 to 497) and Phase – IV (days**

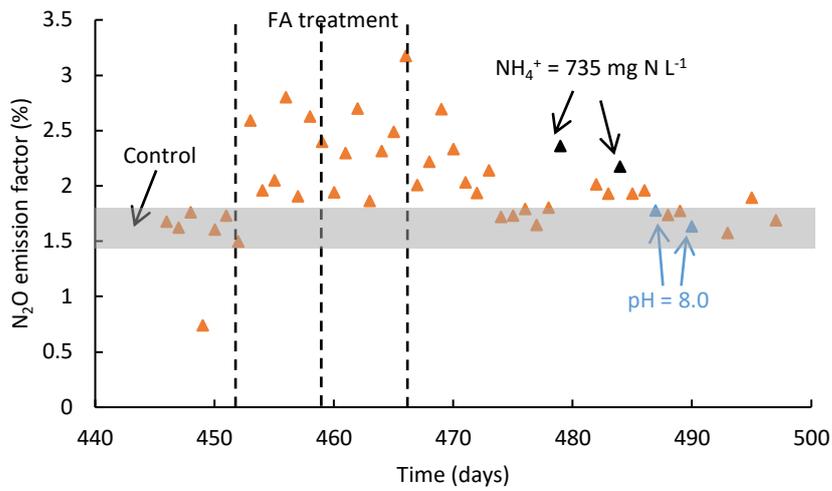
696 **498 to 551). A, volumetric NH<sub>4</sub><sup>+</sup> removal and NO<sub>3</sub><sup>-</sup> production rate; B, relative NO<sub>2</sub><sup>-</sup>**

697 **consumption by AnAOB and NOB, and residual NO<sub>2</sub><sup>-</sup> (in the effluent). The black dashed**

698 **lines indicate FA treatment (30 mg NH<sub>3</sub>-N L<sup>-1</sup>, 3h), whereas FNA treatments (3.0 mg**

699 **HNO<sub>2</sub>-N L<sup>-1</sup>, 3h) are indicated with a green solid line.**

700 **Figure 5**



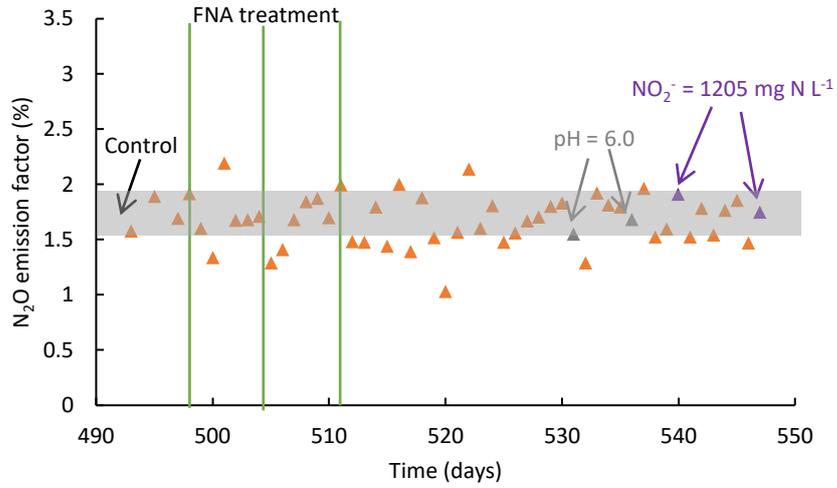
701

702 **Figure 5. Changes in N<sub>2</sub>O emission factors under FA treatment (30 mg NH<sub>3</sub>-N L<sup>-1</sup>, 3h,**

703 **black dashed lines) and associated NH<sub>4</sub><sup>+</sup> (~735 mg N L<sup>-1</sup>, 3h, black triangles) or pH**

704 **shock (~8.0, 3h, blue triangles).**

705 **Figure 6**



706

707 **Figure 6. Changes in N<sub>2</sub>O emission factors under FNA treatment (3 mg HNO<sub>2</sub>-N L<sup>-1</sup>,**

708 **3h, green solid lines) and associated NO<sub>2</sub><sup>-</sup> (~1205 mg N L<sup>-1</sup>, 3h, purple triangles) or pH**

709 **shock (~6.0, 3h, grey triangles).**