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1 **Enrichment and adaptation yield high anammox conversion**
2 **rates under low temperatures**

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Abstract

17 This study compared two anammox sequencing batch reactors (SBR) for one
18 year. SBR_{constantT} was kept at 30°C while temperature in SBR_{loweringT} was
19 decreased step-wise from 30°C to 20°C and 15°C followed by over 140 days at
20 12.5°C and 10°C. High retention of anammox bacteria (AnAOB) and
21 minimization of competition with AnAOB were key. 5-L anoxic reactors with the
22 same inoculum were fed synthetic influent containing 25.9 mg NH₄⁺-N/L and
23 34.1 mg NO₂⁻-N/L (no COD). Specific ammonium removal rates continuously
24 increased in SBR_{constantT}, reaching 785 mg NH₄⁺-N/gVSS/d, and were
25 maintained in SBR_{loweringT}, reaching 82.2 and 91.8 mg NH₄⁺-N/gVSS/d at 12.5
26 and 10°C respectively. AnAOB enrichment (increasing hzsA and 16S rDNA
27 gene concentrations) and adaptation (shift from *Ca. Brocadia* to *Ca. Kuenenia*
28 in SBR_{loweringT}) contributed to these high rates. Rapidly settling granules
29 developed, with average diameters of 1.2 (SBR_{constantT}) and 1.6 mm
30 (SBR_{loweringT}). Results reinforce the potential of anammox for mainstream
31 applications.

Key words: biological nitrogen removal; shortcut nitrogen removal; cold anammox; microbial community structure

32

1. Introduction

33 Partial nitrification/anammox (PN/A) is an autotrophic shortcut nitrogen removal
34 process in which, after partial nitrification, Anammox bacteria (AnAOB) perform
35 anoxic oxidation of ammonium with the produced nitrite as electron acceptor.
36 This process requires less oxygen, abandons the need for organic carbon, and
37 yields a lower sludge production. It can thus present a more cost-efficient
38 treatment compared to the conventional nitrification/denitrification process
39 (Vlaeminck et al. 2012). Many advancements have been made and by the end
40 of 2013, close to one hundred full scale side-stream anammox installations,
41 usually treating digestate, landfill leachate or reject water, have seen the day
42 (Lackner et al. 2014, Mulder et al. 1995, Siegrist et al. 2008, Strous et al. 1999).
43 The next development goal is to introduce anammox in the mainstream or
44 waterline of sewage treatment plants in order to further improve their efficiency
45 in terms of energy consumption (and hence economics) and greenhouse gas
46 emissions.

47

48 The main challenges for PN/A implementation on pretreated sewage, so-called
49 mainstream PN/A, are associated with the development of robust methods to
50 suppress nitrite oxidizing bacteria (NOB) and promote the growth and activity of
51 AnAOB under relatively low influent nitrogen concentrations (40-80 mg NH_4^+ -
52 N/L) and non-negligible amounts of biodegradable organic carbon (which allow
53 ordinary heterotrophic organisms, OHO, to develop and compete for nitrite
54 through denitrification). Furthermore, and of particular interest for regions with a
55 temperate (or cold) climate, relatively low sewage temperatures (below 15°C

56 down to 10°C, or even below) drastically decrease AnAOB growth rates and
57 activity (Cao et al. 2017, Vlaeminck et al. 2012). Their optimal operating
58 temperature has been reported around 35°C - 40°C (Strous et al., 1999, Isaka
59 et al. 2008).

60

61 Several attempts have been made to understand and model the short term
62 impact of decreasing temperature on the specific ammonia removal rate
63 (SARR) by calculating Arrhenius temperature coefficient (also called θ -value) or
64 Activation Energy (E_a). These studies reported a sharp decrease in anammox
65 activity from 15°C and down (up to 10-fold between 30 and 10°C). Maximum
66 reported SARR values (after conversion from N_2 production to NH_4^+
67 consumption based on the theoretical stoichiometry) at 10°C did not go beyond
68 14 mg NH_4^+ -N/gVSS/d (Lotti et al. 2014c, Dosta et al. 2008). However, short-
69 term batch tests do not include the impact of adaptation and selection on the
70 temperature sensitivity of the biomass, and are therefore not sufficient to assess
71 the full potential of anammox at lower temperatures.

72

73 Several reactor strategies have been tested for anammox at lower temperature,
74 with different types of biomass (flocs, granules or carrier-supported biofilms),
75 and types of wastewater. Adaptation could be demonstrated through a shift in
76 optimal temperature upon quickly or gradually decreasing temperature in
77 Moving Bed Biofilm Reactor (MBBR) (Gilbert et al. 2014, Gustavsson et al.
78 2014) and Sequencing Batch Reactor (SBR) systems (Gilbert et al. 2015, Hu et
79 al. 2013). The formation, enrichment and maintenance of anammox granules at

80 lower temperatures has been reported on both synthetic wastewater at 10°C
81 (Hendrickx et al. 2014, Lotti et al. 2014a) and real wastewater at 16°C (Ma et al.
82 2013) and 10°C (Lotti et al. 2014b). In another study where hybrid sludge (flocs
83 + wall attached biofilm) was formed, decreasing temperature caused the SARR
84 to drop tenfold, but the system's performance remained stable at 12°C (Laureni
85 et al. 2015). Stable P/NA performance was also observed when decreasing the
86 temperature to 15°C in 2 MBBRs, however a sudden drop in temperature
87 caused a significant yet reversible drop in anammox activity (Laureni et al.
88 2016). Interestingly, *Brocadia* was the dominant anammox genus in all of the
89 abovementioned studies throughout the adaptation to and operation at low
90 temperature. It should be mentioned that all those studies at low temperature
91 were performed in the presence of oxygen and/or organic carbon and the
92 temperature decrease was often abrupt (big steps) and/or little time (days to a
93 couple of months) was given to renew the slow growing AnAOB. To our
94 knowledge, no study has been done under pure anoxic autotrophic conditions
95 with granular sludge at low temperature.

96

97 The objective of this study was to reveal the maximum potential of so-called
98 'cold anammox' (for temperatures below 15°C), also monitoring potential
99 change in the microbiological structure and the physical properties of granular
100 sludge. In order to achieve an adapted and enriched anammox community, a
101 long-term reactor experiment was designed in which the microbial competition
102 was limited through using anoxic reactors, avoiding AOB and NOB activity, in
103 the absence of organic carbon, avoiding OHO. To do so, an anammox

104 sequencing batch reactor (SBR) operated at high temperature (30°C) was
105 compared to one subjected to subsequent temperature drops for almost 1 year.
106 The impact of lowering the operational temperature from 30 over 20, 15 and
107 12.5 to 10°C on nitrogen removal performance and factors linked to biomass
108 enrichment/adaptation and morphology was examined.

109 **2 Material and Methods**

110 **2.1 Set-up and operation of the reactors**

111 Two anammox SBR were operated in parallel, composed of identical, airtight,
112 mixed, and jacketed vessels (5L), equipped with pH (H 8481 HD, SI Analytics),
113 temperature and dissolved oxygen (DO) probes (Visiferm™, Hamilton). The
114 SBR mode is comprised of four characteristic phases: (1) filling phase (30
115 minutes, inflow of 2.5L influent, hence a volumetric exchange ratio, VER, of
116 50%), during which oxygen was removed from the liquid through stripping with
117 N₂ gas. For the next 30 minutes, the reactors were flushed with a mixture of N₂
118 and CO₂ gas to ensure anoxic conditions and set the pH back to 7.5 before
119 starting the actual (2) reaction phase with a variable duration (between 4h and
120 11.3h). The length of the reaction phase was shortened (lengthened) to
121 increase (decrease) the ammonium loading rate (ALR) when needed. After the
122 reaction phase, mixing was stopped for 30 minutes during the (3) settling phase
123 before starting the (4) discharge phase where 2.5L of supernatant was removed
124 before starting the next cycle. Those operating conditions led to maintain in the
125 system particles that have a settling velocity higher than 0.29 m/h.

126

127 One reactor, $SBR_{\text{constantT}}$, was kept at the reference temperature of 30°C
128 throughout the entire experiment while in the second reactor, $SBR_{\text{loweringT}}$, a
129 stepwise decrease from 30°C to 20°C, 15°C, 12.5°C and finally 10°C was
130 imposed corresponding to phases I, II, III, IV and V respectively. The decrease
131 of temperature was deliberately slow to let the time for the microbial community
132 to adapt to new conditions. More specifically the temperature pattern was
133 chosen according to the following philosophy:

- 134 - Step by step, each step being maintained for at least 2 months, until the
135 Ammonium Removal Efficiency (ARE) recovered a value of 100%, and the
136 Volumetric Ammonium Removal Rate (VARR), i.e. the anammox conversion
137 rate, reached steady state value;
- 138 - The temperature delta was not conservative, but progressively smaller
139 (10°C, 5°C, 2.5°C, 2.5°C), assuming that the last temperature steps would
140 be more detrimental to the anammox bacteria considering previous
141 literature indicating a higher activation constant at low temperature;
- 142 - The rate of temperature decrease was linear with a slope of 2°C/day,
143 2°C/day, 1°C/day, 0.5°C/day for the last, again expecting that the last
144 temperature step would be more critical for anammox bacteria.

145

146 Finally the decrease from 30 to 10°C took 7 months. Calculations were made in
147 order to evaluate if this time was sufficiently long to allow the renewal of most of
148 the biomass initially contained in the reactor considering the AnAOB biomass
149 yield and imposed nitrogen load. These calculations are given in the discussion.

150

151 The reactors were fed with the same synthetic influent which was stored at 4°C
152 in a 150L tank. The influent contained per liter 168.0 mg of NaNO₂, 98.8 mg of
153 NH₄Cl, 165.0 mg of MgSO₄•7H₂O, 143.8 mg of CaCl₂•2H₂O, 6.3 mg of EDTA-
154 Na₂•2H₂O, 775.4 mg of NaHCO₃, 30.0 mg of KH₂PO₄, and 9.1 mg of
155 FeSO₄•7H₂O. Hence, the total nitrogen content was 60 mg N/L, a typical value
156 for municipal wastewater (Metcalf 2003), made up of ammonium and nitrite in a
157 1/1.32 ratio corresponding to the stoichiometry found by (Strous et al. 1998): 25.9
158 mg N-NH₄⁺/L and 34.1 mg N-NO₂⁻/L. Per liter of influent, 2 mL of a trace
159 element solution was added, which contained per liter 430 mg of ZnSO₄•7H₂O,
160 240 mg of CoCl₂•6H₂O, 1.21g of MnCl₂•4H₂O, 250 mg of CuSO₄•5H₂O, 183 mg
161 of Na₂MoO₄•2H₂O, 274.6 mg of NiSO₄•7H₂O, 50.1 mg of Na₂SeO₃•5H₂O, 60.9
162 mg of Na₂WO₄•2H₂O, 14 mg of H₃BO₃ and 15g of EDTA-Na₂•2H₂O.

163 **2.2 Biomass inoculum mix**

164 To provide a high microbial AnAOB diversity, four different non cold-adapted
165 biomasses from stable, long running systems containing AnAOB and operated
166 at rather high temperature (26-30°C) were mixed. Half of the biomass types
167 originated from smaller lab-scale set-ups treating synthetic influent, the others
168 originated from full or pilot scale systems treating centrate from a sewage
169 sludge digester. Three of the systems were PN/A (aerobically operated), the
170 fourth system was anammox (anoxic). The resulting inoculated biomass
171 concentration was 2.3 g VSS/L.

172 **2.3 Anammox activity and chemical analyses**

173 The actual ARE were determined by measuring outlet concentrations of
174 ammonium roughly every two days. Comparable information was obtained from
175 nitrite measurements.

176

177 The systems being operated in sequencing batch mode kinetics could be
178 followed directly during the reaction phase of the cycles. The *in situ* VARR was
179 measured two to three times per week. Samples of mixed liquor (8mL) were
180 taken throughout the reaction phase of one cycle. The ammonium uptake rate
181 was calculated by linear regression (no substrate limitation for ammonium
182 concentration higher than 5 mgN/L for which a linear decrease was observed).

183 Roughly every one or two months, mixed liquor samples were taken from the
184 reactors to determine total (TSS) and volatile (VSS) suspended solids
185 concentrations. For biomass sampling a protocol was carefully set up to collect
186 representative samples. During sampling, the agitation speed was increased
187 (from 50 to 100 rpm) for maximal homogenization of the reactor liquid (and at
188 least one minute before sampling). Sampling and VSS measurements were
189 done in duplicate, and the sample volume (2x50 mL) was chosen to obtain good
190 reproducibility. For each phase, an average biomass concentration (g VSS/L)
191 was calculated (two to three measurements) together with the average *in situ*
192 VARR during the last two weeks (three to four measurements). From this, the *in*
193 *situ* SARR at the end of each phase was estimated. Standard deviations were
194 calculated for average VARR and VSS and propagated in the error bars,
195 allowing for an objective comparison. Student tests (t-tests) were performed to

196 compare mean SARR values between reactors and between consecutive
197 phases within the same reactor.

198

199 All water samples were centrifuged (4°C, 2 minutes at 2,591xg) and
200 supernatant was filtered through 0.2 µm membrane filters prior to nitrogen
201 compounds analysis. Nitrite and nitrate were quantified using
202 spectrophotometric methods, analyses were done automatically using the
203 SMARTCHEM200 (AMS, Italy). Ammonium concentration was measured
204 according to the spectrophotometric Nessler method (APHA 1992). Suspended
205 solids (TSS and VSS) were measured in duplicate according to Standard
206 Methods 2540D and E (APHA 1992).

207 **2.4 Particle size distribution of the biomass aggregates**

208 The particle size distribution (PSD) was determined after 354 days. Triplicate
209 samples of mixed liquor were taken, and particles were visualized with a WILD
210 M420x1.25 (optical zoom set at x6.4) stereomicroscope, operated using a
211 NIKON DS-U2 control unit combined with NIS-elements F software (v.3.2).
212 Photos were taken using a NIKON DS-Fi1 digital microscope camera. For each
213 triplicate, 50 photos were taken. The three series of photos were combined to
214 give one set of 150 photos per reactor for image analysis.

215

216 The first step of the image treatment consisted of a conversion to greyscale
217 using IrfanView software (v.4.10). Next, using Visilog software (v.6.7), the pixel-
218 scale values were converted by a scaling factor (which was calibrated for the
219 magnification). Each grey-scale image underwent a binarization step after which

220 the surface of each particle was quantified and used to estimate the sphere-
221 equivalent diameter. The used script was made so that particles touching the
222 edge of the photo or with a diameter smaller than 15 μm would not be detected
223 (the detection limit corresponded to a minimal number of 10 pixels in the image
224 analysis script). The obtained data was then treated in Excel with a visual basic
225 macro to obtain the number, surface and volume weighted PSD along with
226 additional information such as mean diameters.

227 **2.5 Microbial community analyses**

228 Triplicate samples of 1.5 mL mixed liquor were taken from each reactor, one
229 was used for microbial community analyses, and the other two were stored as
230 back-up. This was done on average once every two to three weeks with higher
231 sampling density around each transition between phases. After centrifugation
232 (20 min at 19,000xg), supernatant was removed and the remaining biomass
233 was immediately frozen in liquid nitrogen. DNA extraction on one of these
234 samples was performed using the FastDNA spinkit™ from MP biomedical
235 following the provided Fastprep® protocol. The extracted DNA was stored at -
236 20°C until further use.

237 In order to map AnAOB enrichment, qPCR was used to quantify the AnAOB
238 based on phylogenetic primers (16S rRNA) and functional primers (hzsA). Also,
239 to follow up the composition of the overall community, MiSeq Illumina
240 sequencing was performed by the GenoToul Genomics and Transcriptomics
241 facility (GeT-PlaGe, Auzeville, France). Sequencing data was processed using
242 the pipeline FROGS, one of the tools proposed on Galaxy, an open web-based
243 platform for genomic research.

244 3. Results

245 3.1. Reactor performance

246 3.1.1. Start-up of the reactors

247 Both reactors were started with the same seeding sludge and operated
248 identically at 30°C for 62 days to allow for adaptation to the new incubations
249 conditions. During this period, both reactors showed a very comparable
250 evolution both in ARE (Figure 1.b) and VARR, starting out around 48 mg NH₄⁺-
251 N/L/d on day 19 and increasing to around 74.4 mg NH₄⁺-N/L/d on day 54
252 (Figure 1.a). Because of this increase in volumetric activity, the duration of the
253 reaction time was decreased twice resulting in an increase ALR: from 40.8 mg
254 NH₄⁺-N/L/d to 47.0 mg NH₄⁺-N/L/d (on day 24) and later to 55.4 mg NH₄⁺-N/L/d
255 (on day 31). While a biomass concentration of 2.3 g VSS/L was initially added,
256 levels of 0.70 and 0.76 g VSS/L, for SBR_{constantT} and SBR_{loweringT} respectively,
257 were retrieved on day 67. This was caused by washout of the smallest
258 aggregates during the first two months of operation, this is illustrated by the
259 increased loss of VSS in the effluent during phase I.

260 Figure 2 shows the average *in situ* specific ammonium removal rates (SARR) at
261 the end of each operating phase. By the end of the phase I, in which both
262 reactors had been operated identically, they had reached very similar *in situ*
263 SARR (103 and 99 mg NH₄⁺-N/gVSS/d for SBR_{constantT} and SBR_{loweringT}
264 respectively). On day 62, temperature in SBR_{loweringT} was decreased to 20°C,
265 whereas from this point on, operating conditions in SBR_{constantT} were left
266 unchanged for the rest of the experiment (T=30°C, ALR = 55.4 mg NH₄⁺-N/L/d).

267 3.1.2. Activity evolution at constant temperature (30°C)

268 For the control reactor $SBR_{\text{constantT}}$, as shown in Figure 1.a, maximal volumetric
269 activity has continuously increased upon inoculation: at the beginning of period
270 II, it increased slightly from 79.2 mg $\text{NH}_4^+\text{-N/L/d}$ on day 69 to 87.6 mg $\text{NH}_4^+\text{-}$
271 N/L/d on day 103. On this day, a leak causing a slight entrance of oxygen (0.05
272 mg $\text{O}_2\text{/L}$) in the headspace was detected. After this leak was repaired, the
273 activity increased even more rapidly to 129.8 mg $\text{NH}_4^+\text{-N/L/d}$ in only three days.
274 This increase in activity with time in $SBR_{\text{constantT}}$ continued almost linearly during
275 the overall study. Finally maximal VARR reached about six times its initial value
276 up to 336 mg $\text{NH}_4^+\text{/L/d}$ at the end of the experiment. At this high VARR, all of
277 the ammonia and nitrite were consumed during the first 45 minutes of the
278 reaction phase. The VARR was higher than the ALR, translating in a part of the
279 reaction phase in which conditions were endogeneous. The ARE remained
280 close to 100% during the entire running period (Figure 1.b). The average
281 effluent concentrations were 0.6 mg $\text{NH}_4^+\text{-N/L}$ and 0.1 mg $\text{NO}_2^-\text{-N/L}$ over the
282 entire operating period.

283 As VSS decreased (roughly by a factor 1.7) during the experiment, the evolution
284 of specific activity is of particular interest: the *in situ* SARR increased by a factor
285 7.5 between the end of phase I (103 mg $\text{NH}_4^+\text{-N/gVSS/d}$) and the end of phase
286 V (785 mg $\text{NH}_4^+\text{-N/gVSS/d}$). This suggests enrichment and/or an adaptation of
287 the anammox biomass; this point will be addressed in the discussion section.

288

289 3.1.3. Activity evolution at decreasing temperature (30°C to 10°C)

290 As shown in Figure 1.a, each temperature decrease step in $SBR_{\text{loweringT}}$ resulted
291 in a decrease of anammox activity which temporarily affected the removal

292 efficiency. This effect was most pronounced when switching from 30 to 20°C
293 (beginning of phase II) and from 15°C to 12.5°C (beginning of phase IV),
294 resulting in a 64.8% and 76.7% drop in activity respectively. However, partial
295 recovery after the temperature drop from 30°C to 20°C, activity took 6 weeks,
296 significantly more than the 3 days required for recovery after the drop from 15°
297 to 10°C, which was probably the result of an adaptation and/or enrichment of
298 the biomass during the 146 days between these events (see chapter 3.2). The
299 effects of decreasing temperature from 20°C to 15°C and 12.5°C to 10°C were
300 less significant, as activity loss was lower (36.3% and 28.4% respectively) and
301 activity recovered within the month. Considering the activity measured at the
302 end of each phase, maximal VARR decreased from 76.2 to 41.0 mg NH₄⁺-N/L/d
303 for temperature decrease from 30°C to 20°C (phases I and II), but was relatively
304 similar at the end of the next three phases (III, IV, V), reaching 24.0 mg NH₄⁺-
305 N/L/d at the end the study. For this reason, it was possible to maintain the
306 nitrogen load constant during the phases II, III, IV and V without impacting that
307 much the ARE, which remained also relatively high during the study, ranging
308 between 57-100% for SBR_{lowering} (Figure 1.b). The average effluent
309 concentrations were 2.7 mg NH₄⁺-N/L and 3.3 mgNO₂⁻-N/L over the entire
310 operating period.

311

312 In SBR_{loweringT} the VSS concentration also decreased during the experiment,
313 from 0.76 g VSS/L to 0.24 g VSS/L. When temperature was decreased to 20°C,
314 *in situ* SARR dropped by 38% between the ends of phase I and II. During this
315 time, the biomass concentration decreased from 0.76 g VSS/L to 0.56 g VSS/L.

316 For the next 11 weeks, SARR remained rather stable, reaching 65.6 mg NH₄⁺-
317 N/gVSS/d at the end of phase III while biomass concentration decreased from
318 0.56 g VSS/L to 0.39 g VSS/L. Further decreases in temperature affected
319 transiently the SARR but did not negatively impact the final SARR obtained
320 after several weeks at the end of the temperature step. On the contrary SARR
321 increased slightly to 82.2 mg NH₄⁺-N/gVSS/d at the end of phase IV and
322 reached 91.8 mg NH₄⁺-N/gVSS/d at the end of phase V. This is the highest
323 reported SARR at such low temperature. These results demonstrate a very
324 successful adaptation and operation of the granular sludge anammox process
325 at 10°C.

326 **Figure 1**

327 **Figure 2**

328 Student tests (data not shown) clearly showed that the SARR in phase II, III, IV
329 and V were significantly different between both reactors. The tests showed no
330 significant difference between two consecutive phases in the same reactor.

331 **3.2 Biomass aggregate size and microbial community analysis**

332 *3.2.1 Biomass particle size distribution*

333 On day 1, both reactors contained a dark brownish and very heterogeneous
334 hybrid sludge, made up mainly of small flocs and suspended biomass and a
335 small amount of granules, features linked to the individual constituents of the
336 inoculum (Table 1). From size fractionation of each biomass type (through
337 sieving) and their respective contribution to the inoculum mix, it was estimated
338 that that 59% of the aggregates were smaller than 315 µm (data not shown).
339 During the first 5 months of reactor operation, the biomass evolved to a bright
340 red, predominantly granular sludge. During the last period of operation (phase

341 V), even at 10°C, the granules demonstrated very good settling properties due
342 to their size and density: To illustrate this, SVI was determined to be 34 mL/g
343 and 26 mL/g for $SBR_{constantT}$ and $SBR_{loweringT}$ respectively on day 323.
344 Furthermore, most of the solids were settled in less than 10 minutes during the
345 settling phase. This corresponds to settling velocities higher than 0.88 m/h.
346 Analysis of the particle size distribution (PSD) was performed on day 354.

347

348 $SBR_{loweringT}$ displays a broader PSD, indicating more variation in granule size
349 compared to the narrower PSD of $SBR_{constantT}$, showing a rather uniform granule
350 size. Also, the maximum observed diameter is higher at low temperature, this is
351 also reflected in a bigger volume median diameter: $D(0.5) = 1551\mu\text{m}$ for
352 $SBR_{loweringT}$ vs. $D(0.5) 1215\mu\text{m}$ for $SBR_{constantT}$. Some granules around 3.3-
353 4.4mm were detected at 10°C, but not observed at 30°C. These results showed
354 that lowering the temperature was not detrimental to the granulation process,
355 and can even generate bigger granules. This is likely due to the lower AnAOB
356 growth rates at low temperatures, which resulted in stronger granules with a
357 higher density (as illustrated by their lower SVI of 26 mL/g compared to 34 mL/g
358 at 30°C). Consequently, granules were less susceptible to breakage, explaining
359 why larger diameters were observed at lower temperatures.

360 *3.2.2 Evolutions in the microbial community*

361 16S rRNA sequencing showed that, right after reactor start-up (day 10), AnAOB
362 made up 64.2% and 57.7% of the total community in SBR_{ref} and SBR_{lowT}
363 respectively. By the end of phase I (day 60) the AnAOB abundances had
364 increased to 79.2% and 80.5% respectively, enrichment was taking place. At

365 the end phase IV abundances in SBR_{constantT} and SBR_{loweringT} were only slightly
366 higher, 81.2% and 87.2% respectively, indicating an AnAOB enrichment that
367 mainly occurred in phase I. The evolution of the microbial population at genus
368 level was monitored throughout the experiment for both reactors.

369

370 Figure 3 shows the evolution at genus level within the AnAOB community from
371 16S rRNA sequencing data. Both reactors started with the same inoculum,
372 which is reflected in a very similar anammox composition at day 10 with a
373 majority of *Ca. Brocadia* (81%), complemented with minor fractions of *Ca.*
374 *Kuenenia* (8%) and *Ca. Jettenia* (11%).

375

376 As long as both reactors were operated identically (the first 60 days), they
377 showed a comparable evolution in their AnAOB profile: an increase in *Ca.*
378 *Kuenenia* abundance together with a slightly lower decrease in *Ca. Brocadia*
379 abundance. *Ca. Jettenia* abundance dropped dramatically to 1% in both
380 reactors, and, regardless of operational temperature, could not re-establish
381 itself dominantly within the community. The initial changes in the AnAOB
382 community during the first 60 days are likely the result of acclimatization to the
383 imposed reactor operating conditions. Regarding the significant loss of
384 suspended solids observed during the first period, a selection by selective
385 retention during settling was likely to occur. Indeed it was observed that the loss
386 of *Ca. Brocadia* (-15%) and *Ca. Jettenia* (-10%), caused by washout of smaller
387 aggregates, made the relative AnAOB abundance of *Ca. Kuenenia* increase
388 (+25%).

389

390 In SBR_{constantT} at 30°C, there was no further major shift. *Ca. Brocadia* remained
391 dominant, and even increased its fraction during Phase II to V, at the expense
392 of a drop in *Ca. Kuenenia* abundance. The population repartition remained
393 roughly unchanged and *Ca. Brocadia* made up 90% of the AnAOB community
394 on day 293 while *Ca. Kuenenia* represented less than 10%.

395

396 In contrast, when reducing the operating temperature to 20°C in SBR_{lowering} a
397 remarkable evolution in the AnAOB community occurred: the *Ca. Brocadia*
398 abundance dropped quickly (from 73 to 29%) whilst the *Ca. Kuenenia*
399 abundance increased just as quickly (from 26 to 68%). Further operation at
400 lower temperatures (15, 12.5 and finally 10°C) did not significantly alter the
401 dominance with the AnAOB community, which on day 293 was comprised of
402 74% of *Ca. Kuenenia* and 26% *Ca. Brocadia*.

403

Figure 3

404 In parallel, the AnAOB enrichment in the biomass was assessed by qPCR at
405 three moments in time. Results in Figure 4 indicate a considerable AnAOB
406 enrichment occurring in both reactors, as reflected from the increase in both
407 hzsA and 16S rDNA copy levels. Interestingly, in both systems most of the
408 abundance increase occurred in the first half of the reactor operation time, by the
409 end of Phase II, with little further enrichment over the second half. The
410 observed trend was similar for hzsA and 16S rDNA copy levels expressed 'per
411 g of biomass' or 'per ng of DNA'.

412

Figure 4

413 **4. Discussion**

414 **4.1 Enrichment and adaptation favoring high specific activities**

415 Our goal was to investigate the maximum potential of anammox at low
416 temperatures (<15°C). After gradually decreasing from the initial temperature of
417 30°C in SBR_{loweringT}, the activity stabilized at 82.2 and 91.8 mg NH₄⁺-N/gVSS/d
418 at 12.5 and 10°C, respectively. The *in situ* SARR in the reference reactor at
419 30°C (SBR_{constantT}) increased continuously, reaching up to 785 mg NH₄⁺-
420 N/gVSS/d. Compared to literature, these specific removal rates at 10°C are
421 considerably higher than previously reported values from reactor studies at low
422 temperature, as illustrated in Table 1. Explanations for these differences in
423 SARR are likely linked to differences in enrichment and/or adaptation degree,
424 which is the consequence of a set of factors fixed in the experimental strategy.
425 obtained a slightly higher SARR at after 131 days of operation 12°C compared
426 to this study after 63 days at the same temperature. The final SARR obtained in
427 this study after adaptation to 10°C was the same as the SARR obtained by Hu
428 *et al.* at 12°C and significantly higher than the SARR obtained at 10°C without
429 adaptation in that same study (Hu et al. 2013).

430 **Table 1**

431 Maximizing space for AnAOB in the biomass is a key factor in both enrichment
432 and adaptation, and yielded from low competition with AnAOB and high
433 retention of AnAOB. Feeding with an autotrophic influent in anoxic conditions
434 indeed minimized competition with AOB, NOB and OHO, the average ratio of
435 nitrite to ammonium removal was 1.27±0.19 and 1.20±0.11 for SBR_{constantT} and
436 SBR_{loweringT} respectively, which was close to the stoichiometry reported by
437 Strous (Strous et al. 1998). Note that, the systems in other studies contained

438 COD or oxygen (Table 1). Maximizing biomass retention enables the slow(est)
439 growing AnAOB to stay or develop in the system. In our study, this was
440 implemented by imposing a low minimum settling velocity, keeping solids
441 settling at > 0.3 m/h into the system. In both reactors, regardless of operational
442 temperature, the bacteria aggregated in rapidly settling granules, which
443 facilitated biomass retention. Nonetheless, biomass retention was not perfect,
444 and sampling and washout renewed an estimated 70 and 100% of the produced
445 AnAOB biomass in $SBR_{\text{constantT}}$ and SBR_{lowering} respectively, based on the
446 nitrogen loading and theoretical growth yield. Anoxic SRT values of 162 and
447 164 days, for $SBR_{\text{constantT}}$ and $SBR_{\text{loweringT}}$ respectively, were obtained, from the
448 estimated solids balance. In other words, operating the systems for more than
449 two times the SRT should in theory replace more than 85% of the original
450 biomass.

451

452 Enrichment in both systems was witnessed by the increase in relative AnAOB
453 abundance in the biomass (qPCR data; Figure 4) in both reactors. According to
454 the qPCR data, most of the enrichment occurred in the first half of the
455 experiment. In $SBR_{\text{loweringT}}$, adaptation was observed at the level of the AnAOB
456 community, with a major shift in dominant AnAOB genus from *Ca. Brocadia* to
457 *Ca. Kuenenia* (16S rRNA gene sequence amplicons; Figure 3). Even though it
458 was not studied here, physiological adaptations within a given AnAOB species
459 could have occurred. There is for instance evidence that AnAOB are able to
460 alter their lipid membrane composition through 'addition synthesis' as a form of
461 homeoviscous adaptation to changes in *in situ* temperature (Rattray et al.

2010). Such type of adaptation might also explain the shift in temperature optimum for AnAOB without significant changes in the dominant genus reported by other authors (Gilbert et al. 2014, Hu et al. 2013).

In order to assess the impact of long-term enrichment and adaptation on the AnAOB performance, a parallel study was performed to test the immediate effect of exposure to low temperatures on the anammox activity, on the four types of biomass used in the inoculum mix. Based on recalculated values from this batch test, the SARR roughly doubled after 67 days of operation at 30°C (end of Phase I; Table 2). After long term operation at 30°C in SBR_{constantT}, the SARR was 16 times higher than the activity of the inoculum at 30°C. Comparably, the final SARR at 12.5-10°C in SBR_{loweringT} was 8-15 times higher than the recalculated inoculum rates under these respective temperatures. As a consequence the SARR ratios (low/high temperature) were similar in the short and long term tests: on short term, exposure to 12.5 and 10°C yielded respectively 20 and 13% of the SARR at 30°C, while the respective values were 17 and 12% for the long-term reactor test (see Table 2) .

Table 2

4.2 Potential AnAOB genus niche differentiation

The results point towards a *Ca. Kuenenia* preference to colder temperatures (20°C and below) and a *Ca. Brocadia* preference to 30°C whereas *Ca. Jettenia* was outcompeted in both reactors. It is interesting to note that each system nonetheless maintained both genera, so the niche differentiation or competition advantage was insufficiently strong to out-compete the one or the other.

487

488 In literature *Ca. Kuenenia* has been only observed as predominant in studies at
489 relatively high temperature. A complete replacement of *Ca. Brocadia* by *Ca.*
490 *Kuenenia* was observed in an anoxic anammox MBBR at 38°C (van der Star et
491 al. 2008). The authors suggested that nitrite limitation was the main driver for
492 this transition, based on the hypothesis that *Ca. Kuenenia* would be an affinity
493 (K) strategist and *Ca. Brocadia* a growth rate (r) strategist. This hypothesis was
494 challenged recently when the opposite switch was observed: *Ca. Brocadia*
495 replaced *Ca. Kuenenia* as the dominant genus in the biofilm of two IFAS
496 reactors operated between 24 and 29°C. The authors claimed that in this case,
497 the tolerance towards higher and toxic nitrite levels affected the selection more
498 than their affinity for low substrate levels (Zheng et al. 2016). *Ca. Kuenenia* has
499 a higher reported tolerance towards nitrite than *Ca. Brocadia*: inhibition occurs
500 for concentration of 180 mg/L and 70 mg/L respectively (Schmid et al. 2003).
501 Neither of these hypotheses seems applicable to our study, since nitrite
502 concentrations were too low to cause inhibition (maximum 17 mg N/L after
503 feeding) yet were never limiting in any of the reactors (operated with rapid
504 feeding in SBR mode). However, from Phase II onwards, the VARR in
505 $SBR_{\text{constantT}}$ was higher than the VALR, meaning that the AnAOB were subjected
506 to endogenous conditions during the final part of the cycle whereas the AnAOB
507 in SBR_{lowering} knew no endogenous period since the VARR was stabilized
508 around the VALR throughout the entire experiment. This presence or absence
509 of an endogenous period may have also played a role in the niche
510 differentiation between *Ca. Brocadia* to *Ca. Kuenenia*.

511

512 This is the first time a switch from *Ca. Brocadia* to *Ca. Kueneria* is observed at
513 low temperature. Almost every other low temperature study, reported that *Ca.*
514 *Brocadia* was and remained the dominant genus throughout the experiment,
515 suggesting that this genus had a competitive advantage over other anammox
516 genera at low temperature (Gilbert et al. 2015, Hendrickx et al. 2014, Hendrickx
517 et al. 2012, Hu et al. 2013, Laureni et al. 2016, Laureni et al. 2015, Lotti et al.
518 2014a, [Sánchez Guillén et al., 2016](#)). Note that during these studies there was
519 always a presence of oxygen and/or organic carbon. [In the study by Sánchez](#)
520 [Guillén and colleagues the maximum DO was 0.2%, but even at such low](#)
521 [concentration AnAOB activity can be impacted \(observed in this study when a](#)
522 [leak was detected on day 69\). It is interesting to note that in the before](#)
523 [mentioned study, average granules size decreased throughout the reactor](#)
524 [operation. The fact that granule size increased in this study was probably due to](#)
525 [the higher NLR and lower mixing intensity \(the SBR reactor in the other study](#)
526 [was fitted with baffles which most likely increased the shear force on the](#)
527 [granules\).](#) Only one study reported on *Ca. Jettenia* as most dominant AnAOB
528 genus in a PN/A Expanded Granular Sludge Bed (EGSB) reactor operated
529 under low DO conditions (0.8 mg/L) to treating COD containing effluent of an
530 upflow anaerobic fixed bed (UAFB) executing anaerobic digestion (AD) (Gao et
531 al. 2015). It is possible that the absence of inhibition (in the case of oxygen) and
532 of competition/partnerships with other bacteria (AOB, NOB and OHO) played a
533 role in the niche differentiation between *Ca. Brocadia* and *Ca. Kueneria*
534 observed in this study at low temperature. Available data on activation energies

535 (E_a) do not allow to differentiate in temperature sensitivity between the two
536 discussed genera. Two studies working with *Ca. Kueneenia* reported E_a -values
537 of 63 kJ/mol between 10°C-40°C (Dosta et al. 2008) and 93 kJ/mol between
538 6°C and 22°C (Isaka et al. 2008), these values fall within the same range as the
539 ones obtained for *Ca. Brocadia* (Hendrickx et al. 2014, Lotti et al. 2014c).

540

541 Further research should focus on understanding the influence key drivers such
542 as high/low affinity for substrate, sensitivity to low temperature and/or to
543 inhibitors etc. have on the niche differentiation between different AnAOB genera
544 and if/how this affects process kinetics/stoichiometry.

545 **4.3 Towards implementation of partial nitrification/anammox**

546 The high activities obtained at low temperatures in this study reinforces the
547 potential of anammox for mainstream application. A key strategy to obtain
548 enrichment and adaptation was to eliminate competition and maximize retention
549 while applying a step-wise decrease in temperature. **Process design and
550 operation will define what activity level can be reached and should therefore be
551 aimed at maximizing the abundance of adapted AnAOB in the biomass.**

552

553 In the end, the high maximal potential AnAOB activities under low temperatures
554 provide a basis for to conceive new PN/A design solutions and associated
555 operational strategies. If 50% of the activity at 10°C could be maintained in a
556 full-scale process with a solids content of 3 g VSS/L, it would work at 135 g
557 $\text{NH}_4^+\text{-N/m}^3/\text{d}$ which corresponds to a hydraulic retention time of about 8 hours,
558 this is a conventional value for activated sludge treating systems domestic

559 sewage. Given the nitrite feeding and anoxic conditions in this study, a two-
560 stage PN/A approach seems to be suitable to go as close as possible to the full
561 potential of an enriched/adapted community, as it avoids the competition with
562 nitrifiers, aerobic heterotrophs and with part of the anoxic heterotrophs.
563 Selective AnAOB retention and enrichment can also be implemented in a **one-**
564 **stage approach**, by selectively increasing the SRT of AnAOB compared to the
565 SRT of the competitors, like in IFAS PN/A systems (Veuillet et al. 2014) or
566 through the use of a selective screen (Han et al. 2016). Future work should
567 verify whether equally high potential rates can be obtained when making a
568 faster and seasonally realistic transition from higher to lower temperatures. Five
569 year temperature gradient data from a WWTP in Germany showed a
570 temperature decrease from 20°C (summer) to 10°C (winter) at a rate of 2°C per
571 month. During the winter temperature could acutely drop down to 7°C for a
572 short period of time (weeks) (Gilbert et al. 2014). In addition, more research
573 should be dedicated to determine any influence of low temperatures on the
574 growth yield. One study in a PN/A SBR estimated that the growth yield lowered
575 with decreasing temperature (Hu et al. 2013) while in another anammox
576 granular MBBR/SBR study no such impact was observed (Hendrickx et al.
577 2014).

578 **5. Conclusions**

579 **Minimal competition and high AnAOB retention resulted in unprecedented**
580 **removal rates of 82 and 92 mg NH₄⁺-N/g VSS/d at 12.5 and 10°C respectively.**
581 **AnAOB enrichment (indicated by increasing hzsA and 16S rRNA gene**
582 **concentrations) and adaptation at genus level (indicated by a shift from Ca.**

583 *Brocadia* to *Ca. Kueneenia* in SBR_{loweringT}) contributed to this performance. Well
584 settling granules were formed and maintained at both 30°C and 10°C, indicating
585 that lowering temperature is not detrimental to granulation and can even
586 increase granule size. These results provide new insights that reinforce the
587 potential of cold anammox applications for mainstream N-removal.

588

589 E-supplementary data of this work can be found in online version of the paper

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595 **7. References**

596 APHA, A. (1992) WPCF (American Public Health Association, American

597 Waterworks Association, Water Pollution Control Federation)(1992)

598 Standard methods for the examination of water and wastewater. Standard

599 methods for the Examination of Water and Wastewater 17.

600 Cao, Y., van Loosdrecht, M.C.M. and Daigger, G.T. (2017) Mainstream partial

601 nitritation–anammox in municipal wastewater treatment: status, bottlenecks,

602 and further studies. *Applied Microbiology and Biotechnology* 101, 1365-

603 1383.

604 Dosta, J., Fernandez, I., Vazquez-Padin, J.R., Mosquera-Corral, A., Campos,

605 J.L., Mata-Alvarez, J. and Mendez, R. (2008) Short- and long-term effects of

606 temperature on the Anammox process. *Journal of Hazardous Materials* 154,
607 688-693.

608 Gao, D.-W., Huang, X.-L., Tao, Y., Cong, Y. and Wang, X.-I. (2015) Sewage
609 treatment by an UAFB–EGSB biosystem with energy recovery and
610 autotrophic nitrogen removal under different temperatures. *Bioresource
611 Technology* 181, 26-31.

612 Gilbert, E.M., Agrawal, S., Karst, S.M., Horn, H., Nielsen, P.H. and Lackner, S.
613 (2014) Low Temperature Partial Nitritation/Anammox in a Moving Bed
614 Biofilm Reactor Treating Low Strength Wastewater. *Environmental Science
615 & Technology* 48, 8784-8792.

616 Gilbert, E.M., Agrawal, S., Schwartz, T., Horn, H. and Lackner, S. (2015)
617 Comparing different reactor configurations for Partial Nitritation/Anammox at
618 low temperatures. *Water Research* 81, 92-100.

619 Gustavsson, D., Persson, F. and la Cour Jansen, J. (2014) Manammox–
620 mainstream anammox at Sjölanda WWTP. Proceedings from the IWA World
621 Water Congress and Exhibition, September 21-26, Lisbon, Portugal (2014)

622 Han, M., Vlaeminck, S.E., Al-Omari, A., Wett, B., Bott, C., Murthy, S. and De
623 Clippeleir, H. (2016) Uncoupling the solids retention times of flocs and
624 granules in mainstream deammonification: A screen as effective out-
625 selection tool for nitrite oxidizing bacteria. *Bioresource Technology* 221,
626 195-204.

627 Hendrickx, T.L.G., Kampman, C., Zeeman, G., Temmink, H., Hu, Z., Kartal, B.
628 and Buisman, C.J.N. (2014) High specific activity for anammox bacteria

629 enriched from activated sludge at 10 degrees C. *Bioresource Technology*
630 163, 214-221.

631 Hendrickx, T.L.G., Wang, Y., Kampman, C., Zeeman, G., Temmink, H. and
632 Buisman, C.J.N. (2012) Autotrophic nitrogen removal from low strength
633 waste water at low temperature. *Water Research* 46, 2187-2193.

634 Hu, Z., Lotti, T., de Kreuk, M., Kleerebezem, R., van Loosdrecht, M., Kruit, J.,
635 Jetten, M.S. and Kartal, B. (2013) Nitrogen removal by a nitritation-
636 anammox bioreactor at low temperature. *Applied and environmental*
637 *microbiology* 79, 2807-2812.

638 Isaka, K., Date, Y., Kimura, Y., Sumino, T. and Tsuneda, S. (2008) Nitrogen
639 removal performance using anaerobic ammonium oxidation at low
640 temperatures. *Fems Microbiology Letters* 282, 32-38.

641 Lackner, S., Gilbert, E.M., Vlaeminck, S.E., Joss, A., Horn, H. and van
642 Loosdrecht, M.C.M. (2014) Full-scale partial nitritation/anammox
643 experiences - An application survey. *Water Research* 55, 292-303.

644 Laurenzi, M., Falås, P., Robin, O., Wick, A., Weissbrodt, D.G., Nielsen, J.L.,
645 Ternes, T.A., Morgenroth, E. and Joss, A. (2016) Mainstream partial
646 nitritation and anammox: long-term process stability and effluent quality at
647 low temperatures. *Water Research* 101, 628-639.

648 Laurenzi, M., Weissbrodt, D.G., Szivák, I., Robin, O., Nielsen, J.L., Morgenroth,
649 E. and Joss, A. (2015) Activity and growth of anammox biomass on
650 aerobically pre-treated municipal wastewater. *Water Research* 80, 325-336.

651 Lotti, T., Kleerebezem, R., Hu, Z., Kartal, B., Jetten, M. and van Loosdrecht, M.
652 (2014a) Simultaneous partial nitrification and anammox at low temperature
653 with granular sludge. *Water Research* 66, 111-121.

654 Lotti, T., Kleerebezem, R., van Erp Taalman Kip, C., Hendrickx, T., Kruit, J. and
655 Van Loosdrecht, M. (2014b) Anammox growth on pretreated municipal
656 wastewater. *Environmental science & technology* 48,7874-80.

657 Lotti, T., Kleerebezem, R. and van Loosdrecht, M. (2014c) Effect of temperature
658 change on anammox activity. *Biotechnology and bioengineering* 112, 98-
659 103.

660 Ma, B., Peng, Y., Zhang, S., Wang, J., Gan, Y., Chang, J., Wang, S., Wang, S.
661 and Zhu, G. (2013) Performance of anammox UASB reactor treating low
662 strength wastewater under moderate and low temperatures. *Bioresource*
663 *Technology* 129, 606-611.

664 Metcalf, E. (2003) *Waste water engineering: treatment and reuse*, 4th edn.
665 Revised by Tchobanoglous G, Burton FL, Stensel HD, McGraw-Hill, New
666 York.

667 Mulder, A., van de Graaf, A.A., Robertson, L.A. and Kuenen, J.G. (1995)
668 Anaerobic ammonium oxidation discovered in a denitrifying fluidized bed
669 reactor. *FEMS Microbiology Ecology* 16, 177-183.

670 Rattray, J.E., van de Vossenberg, J., Jaeschke, A., Hopmans, E.C., Wakeham,
671 S.G., Lavik, G., Kuypers, M.M., Strous, M., Jetten, M.S. and Schouten, S.
672 (2010) Impact of temperature on ladderane lipid distribution in anammox
673 bacteria. *Applied and environmental microbiology* 76, 1596-1603.

674 Sánchez Guillén, J.A., Lopez Vazquez, C.M., de Oliveira Cruz, L.M., Brdjanovic,
675 D., van Lier, J.B. 2016. Long-term performance of the Anammox process
676 under low nitrogen sludge loading rate and moderate to low temperature.
677 *Biochemical Engineering Journal*, 110, 95-106.

678 Schmid, M., Walsh, K., Webb, R., Rijpstra, W.I., van de Pas-Schoonen, K.,
679 Verbruggen, M.J., Hill, T., Moffett, B., Fuerst, J., Schouten, S., Sinninghe
680 Damsté, J.S., Harris, J., Shaw, P., Jetten, M. and Strous, M. (2003)
681 Candidatus "*Scalindua brodae*", sp. nov., Candidatus "*Scalindua wagneri*",
682 sp. nov., Two New Species of Anaerobic Ammonium Oxidizing Bacteria.
683 *Systematic and Applied Microbiology* 26, 529-538.

684 Siegrist, H., Salzgeber, D., Eugster, J. and Joss, A. (2008) Anammox brings
685 WWTP closer to energy autarky due to increased biogas production and
686 reduced aeration energy for N-removal. *Water Science and Technology* 57,
687 383-388.

688 Strous, M., Heijnen, J.J., Kuenen, J.G. and Jetten, M.S.M. (1998) The
689 sequencing batch reactor as a powerful tool for the study of slowly growing
690 anaerobic ammonium-oxidizing microorganisms. *Applied Microbiology and*
691 *Biotechnology* 50, 589-596.

692 Strous, M., Kuenen, J.G. and Jetten, M.S. (1999) Key physiology of anaerobic
693 ammonium oxidation. *Applied and environmental microbiology* 65, 3248-
694 3250.

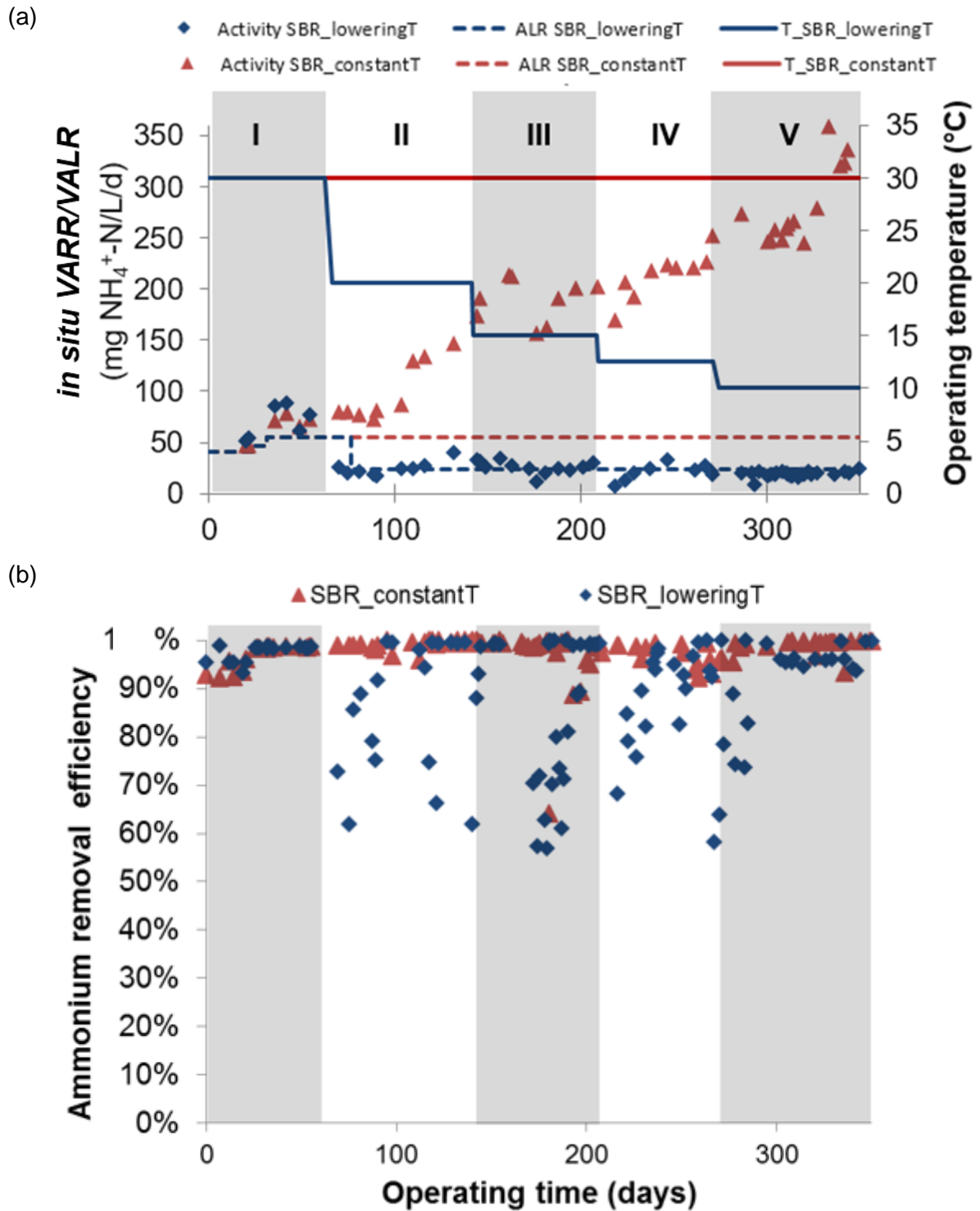
695 van der Star, W.R.L., Miclea, A.I., van Dongen, U.G.J.M., Muyzer, G.,
696 Picioreanu, C. and van Loosdrecht, M.C.M. (2008) The membrane

697 bioreactor: A novel tool to grow anammox bacteria as free cells.
698 Biotechnology and Bioengineering 101, 286-294.

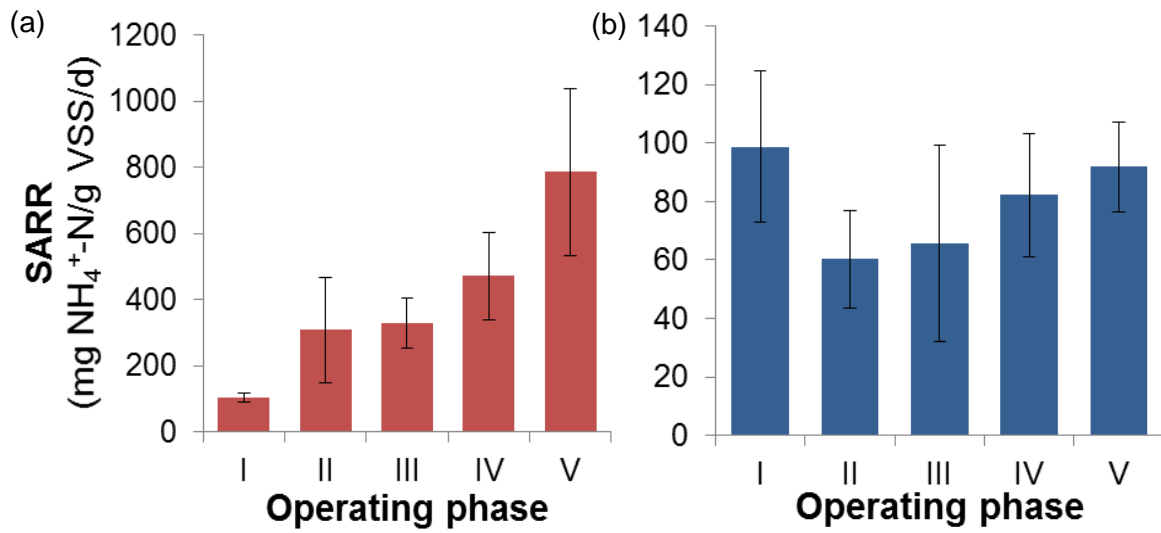
699 Veuillet, F., Lacroix, S., Bausseron, A., Gonidec, E., Ochoa, J., Christensson,
700 M. and Lemaire, R. (2014) Integrated fixed-film activated sludge ANITA™
701 Mox process—a new perspective for advanced nitrogen removal. Water
702 Science & Technology 69, 915-922.

703 Vlaeminck, S.E., De Clippeleir, H. and Verstraete, W. (2012) Microbial resource
704 management of one-stage partial nitritation/anammox. Microbial
705 Biotechnology 5, 433-448.

706 Zheng, B., Zhang, L., Guo, J., Zhang, S., Yang, A. and Peng, Y. (2016)
707 Suspended sludge and biofilm shaped different anammox communities in
708 two pilot-scale one-stage anammox reactors. Bioresource Technology 211,
709 273-279.



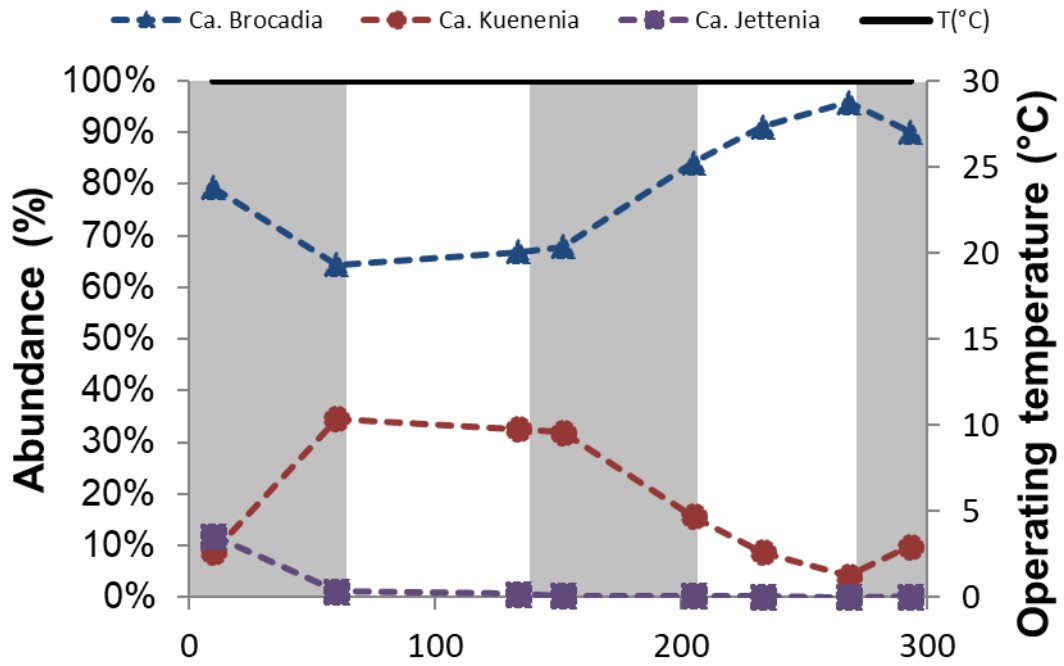
710 **Figure 1** – Evolution of the operating temperature, *in situ* volumetric ammonium removal rate and
 711 volumetric ammonium loading rate (a) and ammonium removal efficiency (b) in SBR_{constantT} (red) and
 712 SBR_{loweringT} (blue).
 713



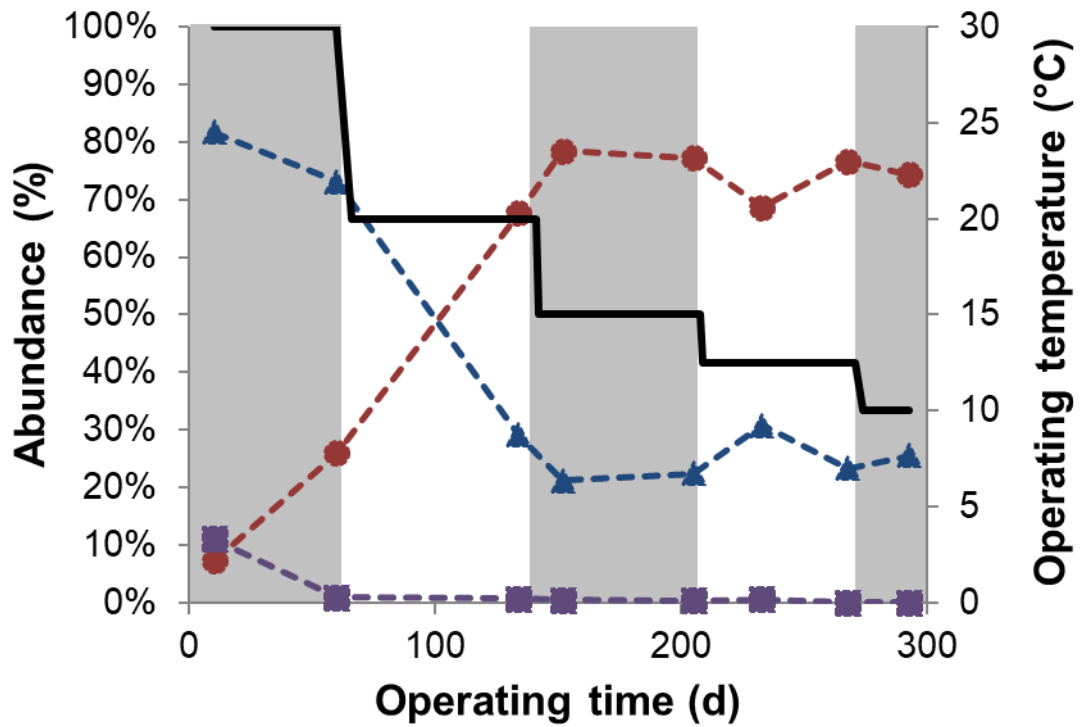
714 **Figure 2** – Average *in situ* specific ammonium removal rates (SARR) at the end of each operating phase
 715 for SBR_{constantT} (a) and SBR_{loweringT} (b). The corresponding days at the end of each phase are 62 (I), 141
 716 (II), 208 (III), 271 (IV) and 349 (V). Error bars represent standard deviations (n≥3)

717

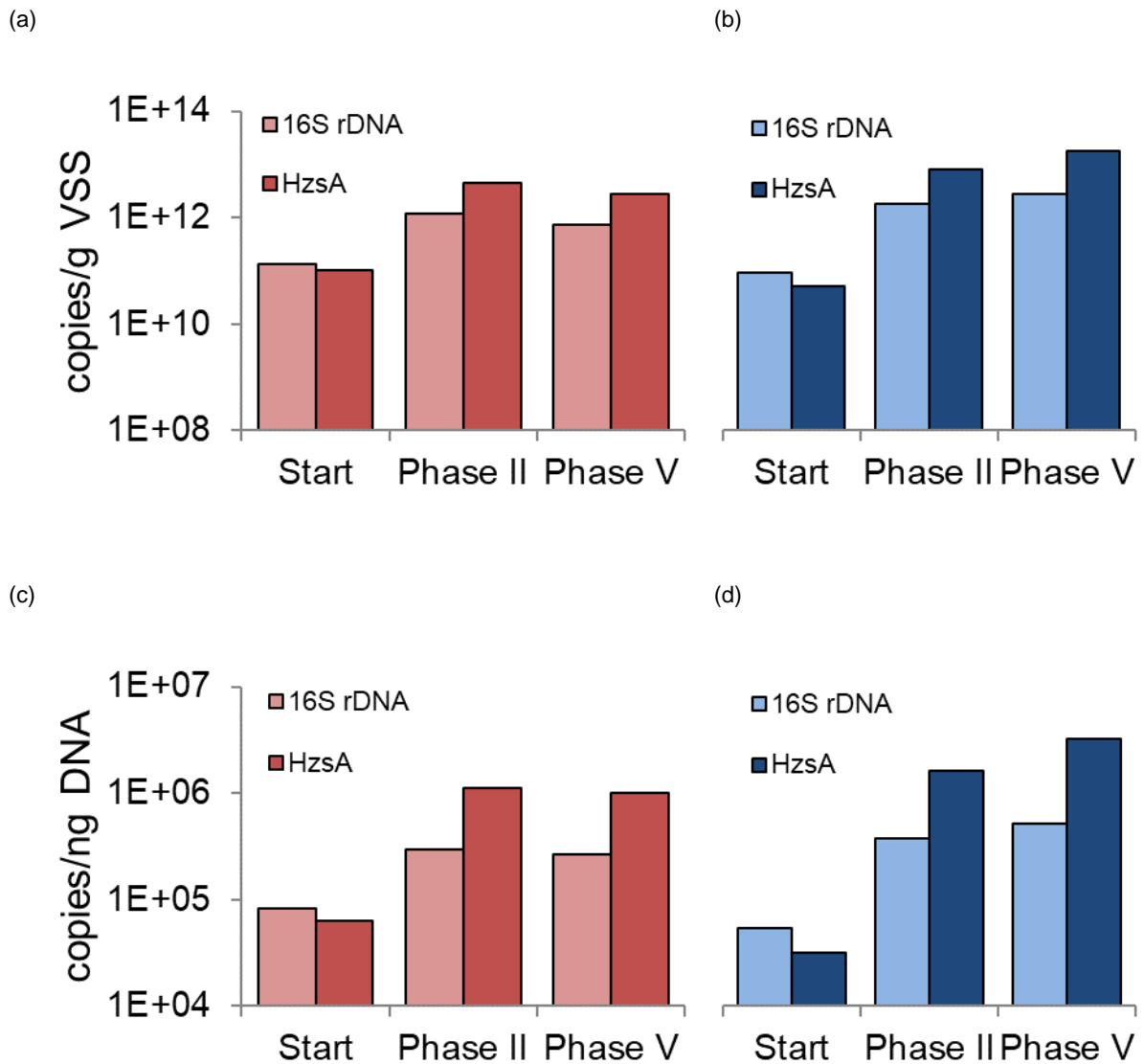
(a)



(b)



718 **Figure 3** – Evolutions within the AnAOB community for $SBR_{constantT}$ (a) and $SBR_{loweringT}$ (b): relative
719 abundances of *Ca. Brocadia*, *Ca. Kuenenia* and *Ca. Jettenia* genera, along with the operating
720 temperature.



721 **Figure 4** - Evolution of overall AnAOB abundance as determined by qPCR analyses based on AnAOB
 722 16S rDNA (lighter bars) and the HzsA functional gene (darker bars). Top row shows the data expressed
 723 per g of VSS for SBRconstantT (a) and SBRLoweringT (b). Bottom rows show the same data expressed
 724 per ng of DNA for SBRconstantT (c) and SBRLoweringT (d). The corresponding sampling times for Start,
 725 Phase II and Phase V are 10, 134 and 293 days, respectively.

726 **Table 1** - Specific ammonium removal rates (SARR, expressed in mg NH₄⁺-N/g VSS/d) and selected
 727 influencing factors for anammox based processes at low temperatures (<15°C) and in different reactor
 728 configurations. Moving Bed Biofilm Reactor (MBRR), Sequencing Batch Reactor (SBR), Partial
 729 Nitritation/Anammox (PN/A), Chemical oxygen demand (COD)

Process configuration; Process type	Substrates for competitors	T _{low} (°C)	Period lowering T (d)	Period at T _{low} (d)	SARR	Reference
Granular MBBR/SBR; Anammox	COD	10	-	722	13-19	Hendrickx <i>et al.</i> 2014
SBR; PN/A	O ₂	12	10 (25-12°C)	131	93	Hu <i>et al.</i> 2013
Granular upflow fluidized bed; Anammox	COD	10	127 (20-10°C)	151	35	Lotti <i>et al.</i> 2014b
Granular SBR; Anammox	O ₂	13.2	523 (30.5-13.2°C)	75	15.1	Sánchez Guillén <i>et al.</i> , 2016
Granular SBR; Anammox	-	12.5	208 (30-12.5°C)	63	82	This study
		10	257 (30-10°C)	78	92	

730

731 **Table 2** – Comparison of the short vs. long term effect of temperature decrease on SARR. Between
 732 brackets the Phase number from the long-term reactor experiment is given. For the short term test,
 733 biomass was directly exposed to lower temperatures and the data from individual biomass tests were used
 734 to calculate a weighted average, based on the different biomass contributions in the inoculum of the
 735 reactor test.

	Short term experiment	Long term experiment	
	Inoculum mix	SBR _{constantT}	SBR _{loweringT}
SARR 30°C (mg NH ₄ ⁺ -N/gVSS/d)	49.2	104 (I); 471 (IV); 785 (V)	98.7 (I)
SARR 12.5°C (mg NH ₄ ⁺ -N/gVSS/d)	9.99	-	82.2 (IV)
SARR 10°C (mg NH ₄ ⁺ -N/gVSS/d)	6.30	-	91.8 (V)
SARR ratio 12.5°C/30°C (%)	20	-	17*
SARR ratio 10°C/30°C (%)	13	-	12**

736 * ratio SBR_{loweringT}/SBR_{constantT} for Phase IV

737 ** ratio SBR_{loweringT}/SBR_{constantT} for Phase V

Electronic Annex

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